

Imperial College

OF SCIENCE, TECHNOLOGY AND MEDICINE

Effects of NO_x and NH_3 on lichen communities and urban ecosystems

A Pilot Study

A report produced by Imperial College & The Natural History Museum, as partners in the

A.P.R.I.L.

Network for the Department for Environment, Food and Rural Affairs

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A.P.R.I.L. AIR POLLUTION RESEARCH IN LONDON A research network supported by EPSRC, DEFRA & EA

APRIL is a multidisciplinary research network that aims to deliver research projects identified by academics, local and national government and other stakeholders working in urban environments. Through regular seminars, workshops and meetings a large research programme has arisen covering the following major topics: Modelling, Measurements, Meteorology, Natural Environment, Health, Planning, Economic & Social Issues, Transport and Indoor Air Quality. A key element of the research programme is collaboration between many different disciplines and organisations in London and other parts of the UK. By combining elements of each specialist group's research programme two large consortium proposals have arisen and several more are in preparation. The first, Meteorology and Air Pollution in London's Environment (MAPLE, measures meteorological and pollution parameters of the air mass coming into London, in and above the city itself and the air mass leaving the city. It involved climatic and pollutant measurements at three sites. Cliffe on the southeastern perimeter of the city, Regents Park in the centre of London and Silwood Park to the West. Campaigns to assess pollution at ground level and in the vertical profile are proposed to provide a greater understanding of the sources of particulates and oxides and nitrogen and their chemistry and dispersion over a large city to specifically inform decisions on the most appropriate abatement strategies to meet London's air quality objectives and generally to improve scientific understanding of the urban environment. Funding is required.

The second consortium proposal, Dispersion of Air Pollution & Penetration into the Local Environment (DAPPLE), funded by EPSRC, investigates the finer scale pollution problems at street corners and junctions where the highest concentrations from traffic emissions arise and where human exposure is potentially greatest. This study is located in the Marylebone Road, Westminster and combines expertise from five universities. It will provide much needed measurements and lead to improvements in the modelling tools used to assess the impact of pollutants on health, including the accidental or terrorist releases of toxic or flammable gases, spatial design and the location of buildings. Other funded projects are studying the impact of pollution on plants and birds in the city and a major health study is currently in preparation.

APRIL host seminars and conferences, specialist workshops and meetings, and members participate at a national and international level at air quality events. The Network, established in 1999, is led by a steering committee chaired by Professor Helen ApSimon of Imperial College, London, supported by: Professor Mike Batty, UCL, Professor Bernard Fisher (Environment Agency), Professor Frank Kelly (King's College), Professor Alan Robins (Surrey), Professor Lord Julian Hunt (UCL), Dr. Roy Colvile (Imperial), Dr. Steve Smith (King's), Dr. Claire Burton (EPSRC), Dr. Janet Dixon (DEFRA), Jim Storey (Environment Agency (EA), David Hutchinson (GLA), Chris Lee (ALG), Steve Hedley (King's College), Linda Davies (Network Co-ordinator, Imperial College). http://www.airpollution.org.uk Linda.davies@ic.ac.uk

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Executive Summary

The increase of oxidised nitrogen in urban areas and reduced nitrogen in rural areas has been widely recognised in Europe. Protocols using lichens have been developed to monitor ammonia pollution in the Netherlands using nitrophyte and acidophyte indicator species (van Herk) and an EU recording method to detect environmental changes. This small pilot study tests the application of these protocols in the UK in selected urban and rural sites. A complimentary study using lichens transplanted from a rural site to the centre of London was carried out to assess eco-physiological changes in selected species.

- The EU Directional Quantitative Lichen Monitoring protocol was used to detect changes in epiphytic lichen diversity on trunks of ash trees between sites in inner and outer London.
- Twigs with a healthy lichen community were transplanted within a clean air site in Somerset and to the wildlife garden at the Natural History Museum adjacent to Cromwell Rd in the vicinity of NOx monitoring guages. The health of individual thalli of selected species was assessed using chlorophyll fluorescence.
- The van Herk method was tested on oak trees in rural localities in different climatic regions in SW (North Wyke, Devon) and E England (Thetford) in the vicinity of ammonia monitoring networks and Sites of Special Scientific Interest.

The pilot survey in London showed differences in lichen diversity on ash trees between inner and outer London sites correlating with concentrations of transport emissions. However the high diversity on oak trees in central London suggests that species recovery is not as limited by slow dispersal mechanisms as previously considered. Over a 3-month period transplanted material from Somerset to a roadside site across a NOx gradient in London showed great variation in persistence and vitality between species. The chlorophyll fluorescence data requires further testing and evaluation over a longer time period to develop its application as a biomarker of stress for NOx.

Indices of nitrophyte and acidophyte species were correlated with ammonia levels (Thetford) and distance from source (North Wyke). However, many nitrophyte species defined in Holland were absent from the sites in the UK, suggesting that indicator species must be evaluated on a regional basis. The importance of both climate and pollution history in the UK was also highlighted by the survey. Highly pollution sensitive Lobarion species that are also indicators of ecological continuity were present at North Wyke whilst at Thetford acidophytes on ancient trunks in conservation sites testified to former acidification. In both sites younger trees and particularly twigs supported the majority of nitrophyte species.

The widespread occurrence of nitrophytes in urban and rural sites investigated in the pilot project suggests that both oxidised and reduced nitrogen is readily assimilated by these lichens. Further work is needed to define species that can be used as indicators of atmospheric nitrogen in its various forms across Britain. It is to be expected that species new to Britain or science will be discovered as was found during surveys in the Netherlands.

This pilot study has highlighted the need for standardised recording techniques for epiphytic lichens in combination with physico-chemical data. This is necessary to develop practical indicator scales appropriate to the UK that can be used to monitor the new pollution climate.

Introduction

This pilot study investigates the impact of reduced and oxidised nitrogen on sensitive vegetation at selected sites in England. The Review and Assessment of Air Quality (DETR, 2000) resulted in Air Quality Management Areas (DEFRA, 2000) being declared by twenty-nine of the thirty-three local authorities in London making it the largest urban area in the UK in breach of the Health Objectives for nitrogen dioxide of 21 ppb (annual mean). Objectives for total oxides of nitrogen (16 ppb) and sulphur dioxide (8 ppb) have been determined to protect sensitive vegetation and ecosystems from harm and although not applicable in urban environments, nevertheless are relevant.

Data collected by Clapp (Clapp & Jenkin 2001^{*}) of measurements of total NOx at the roadside in the centre of Westminster averaged 210 ppb NOx, falling to 73 and 23 ppb respectively at the outer London background sites of Hillingdon and Teddington. There are huge diurnal and seasonal variations in values. The modelling of nitrogen dioxide concentrations by the Greater London Authority (GLA, 2001) reproduced in Section 1, Figure 1.1 clearly demonstrates the gradient in nitrogen dioxide concentrations across London and was used to select three inner London sites and three outer London sites where the diversity, frequency and vitality of lichens and bryophytes were recorded on a single phorophyte, ash (*Fraxinus excelsior*). Lichen diversity on oak (*Quercus spp*) at one central location was also recorded. More detailed investigations at the microscale level were carried out on transplanted lichen material in central London at the Natural History Museum wildlife garden (Cromwell Road).

Background concentrations of nitrogen dioxide decline along a transect from central London through the suburbs and into Surrey where only three local authorities needed to declare Air Quality Management Areas. Heathland communities are particularly sensitive to nitrogen input and formed the basis of a second study looking at nitrogen accumulation in *Calluna vulgaris* and the use of isotopic signatures as an index of traffic-derived pollution in this species (Power & Collins, in prep.).

The primary emission sources of nitrogen in London are transport and heating, contrasting sharply with rural areas where reduced nitrogen from farming activities is frequently the most common form of atmospheric pollution. Intensive agricultural activity in the form of a poultry unit in Norfolk and a dairy farm in Devon provide the third study area where the influence of ammonia on bark pH and lichen communities as determined by van Herk (1999, 2001) was assessed.

The data from this pilot study offer an insight into temporal and spatial differences in selected sensitive species under changing environmental conditions. The project does not

^{*} Daily daylight-averaged NOx, annual mean concentration using 1998-1999 data ppbv.

consider other primary or secondary pollutants from these sources, climate change or differences in management practice.

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- 1. The Air Quality Strategy for England, Scotland, Wales and Northern Ireland (2000). Department of Environment, Transport and the Region. HMSO, London.
- 2. The Air Quality Regulations (2000). Department for the Environment, Food and Rural Affairs.
- 3. Clapp, L.J. & Jenkin, M.E. (2001). Analysis of the relationship between ambient levels of O₃, NO₂ and NO as a function of NOx in the UK. *Atmospheric Environment* **35**: 6391-6405.
- 4. The Mayor's Air Quality Strategy. A consultation document. (2001). Modelled Concentrations of Nitrogen Dioxide, 1999. Greater London Authority (GLA), Romney House, Marsham Street, London.
- 5. Van Herk, C.M. (1999). Mapping of ammonia pollution with epiphytic lichens in the Netherlands. *Lichenologist* **31**: 9-20.
- 6. Van Herk, C.M. (2001) Bark pH and susceptibility to toxic air pollutants as independent causes of changes in epiphytic lichen composition in space and time. *Lichenologist* **33**: 419-441.

This report has been compiled in two sections corresponding to the 2 separate projects funded within the *Terrestrial Umbrella – Eutrophication and Acidification of Terrestrial Ecosystem in the UK* Programme to Imperial College and The Natural History Museum by the Centre of Ecology and Hydrology, Monks Wood. A third section '*Impacts of urban NO_x pollution on vegetation in London*' by Sally Power and Tilly Collins (Imperial College) is coming out as a separate publication.

Section 1. Impacts of NO_x pollution on lichens

Part 1. Corticolous Lichens in London

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Part 2. Investigating the impact of NO_x on transplants

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Section 2. Assessing the role of biological monitoring using lichens to map excessive ammonia (NH_3) deposition in the UK.

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SECTION 1

Impacts of NO_x Pollution on Lichens

PART 1: CORTICOLOUS LICHENS IN LONDON

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1. INTRODUCTION

This pilot study investigates the diversity and distribution of corticolous lichens on a single phorophyte, ash (*Fraxinus excelsior*) at six sites in London and on oak (*Quercus*) at one site. It was proposed as part of a wider study covering all thirty-two London Boroughs and the City of London recording corticolous lichens, bryophytes and fungi on five *Fraxinus* at each site and young oak (*Quercus*) at selected sites.

The project investigates both spatial and temporal trends in relation to species diversity, vitality and community structure, making reference to the EU Limit values (Air quality framework Directive, 1996) to protect sensitive species from the effects of sulphur dioxide and oxides of nitrogen. The annual mean of 16 ppb ($30 \mu g/m^3$) determined for oxides of nitrogen (NOx) is widely exceeded in London. In contrast the sulphur dioxide annual mean and winter mean (1 Oct to 31 Mar) of 8 ppb ($20 \mu g/m^3$) are rarely exceeded. These values have been incorporated into the Air Quality (England) Regulations 2000 (DETR, 2000). They do not apply in urban environments but nevertheless are critical levels for the protection of sensitive species. Lichens and bryophytes are less well protected from atmospheric pollutants than higher plants because they do not possess cuticles or stomata. Some species, however, are better adapted and demonstrate greater tolerance than others. Only one corticolous species, *Lecanora dispersa* was recorded in central London in 1967 (Laundon, 1970) at a time when the limited diversity in most urban environments was attributed to the impact of sulphur dioxide.

This study applies, for the first time in the UK, the proposed EU Quantitative Lichen Monitoring protocol proposed as a standardised recording technique to assess lichen diversity as an indicator of environmental change.

For this pilot study Westminster, Southwark and Tower Hamlets were selected as the inner London sites, all within 6 km of Charing Cross, and outer London locations were selected in Enfield, Harrow and Bromley. A contour map prepared by the Greater London Authority demonstrating the concentration gradient of nitrogen dioxide decreasing with distance from central London to the suburbs is provided (Appendix B).

2. SITE DESCRIPTIONS

2.1 Inner London

2.1.1 Westminster Regent's Park (TQ 282 833) Distance from Charing Cross 3 km

Regent's Park evolved from a 16th Century Royal Hunting Estate into a public park in the early 1800s. Huge areas of trees had been removed to meet timber demands, creating a major challenge in landscaping and design on an area overlying a great depth of London clay. An ambitious tree planting programme, formal gardens and ornamental lakes have transformed the park into a major recreational centre with a wide variety of flora and fauna covering a total area of over 170 hectares.

Ash trees in the vicinity of the York Bridge entrance on the Outer Ring Road, close to the Marylebone Road were selected for the survey, and young oaks adjacent to the Heather Garden. The oaks are situated further from the Inner Road, away from traffic influences and are sheltered by a children's play area. They are closer to London Zoo, and adjacent to the Bird House and lake.

Air pollution data from the kerbside Marylebone monitoring station 0.5 km from York Bridge is provided below but background estimated concentrations calculated by Westminster Local Authority using 1997 data suggest that lowest concentrations of nitrogen dioxide at the present time would be in the region of 31 ppb. The range of total NOx measured at this site is between 50 and 500 ppb (Clapp & Jenkin, 2001).

| Annual Average NO ₂ | 1997 | Background | 31 ppb |
|-----------------------------------|------|------------|---------|
| Annual Average NO ₂ | 1999 | Kerbside | 47 ppb |
| Annual hourly max NO ₂ | 1999 | Kerbside | 169 ppb |
| Annual Average SO ₂ | 1999 | Kerbside | 5 ppb |

Nitrogen dioxide measurements for all sites are taken from the SEIPH 1999 Annual Report (SEIPH, 2001), unless otherwise stated, and represent measurements recorded by the London Air Quality Network.

2.1.2 Southwark Burgess Park (TQ 333 778) Distance from Charing Cross 4 km

This recreational Park was created in the post war period for the enjoyment of the people of Southwark and includes several ornamental lakes, play areas and landscape features with a wide variety of trees, including lime, plane, and ash, many shrubs and cultivated beds. It covers an area of approximately thirty-five hectares and lies between two major road links. Background concentrations of nitrogen dioxide measured in 1997 with diffusion tubes give an average annual value of 21 ppb in the centre of the park (Southwark, 2001):

| Annual Average NO ₂ | 1999 Roadside | 39 ppb |
|-----------------------------------|-----------------|---------|
| Annual hourly Max NO ₂ | 1999 Roadside | 112 ppb |
| Annual Average SO ₂ | 1999 Background | 4 ppb |

SO₂ values are taken from the DEFRA website maintained by AEA/NETCEN (AEA, 2001).

2.1.3 Tower Hamlets Victoria Park (TQ 353 835) Distance from Charing Cross 6 km

This park was created in 1888 in response to the demands of local residents for recreational space in East London. It extends across some 35 hectares and is beautifully landscaped with a variety of mixed broadleaf trees, shrubs, ornamental beds and two lakes. The park is bordered on both sides by major transport links and modelled data (Tower Hamlets, 2002) suggest that background concentrations of nitrogen dioxide remain at approximately 26 ppb in the surveyed areas.

| Annual Average NO ₂ | 2002 Background | 26 ppb |
|-----------------------------------|-----------------|---------|
| Annual Average NO ₂ | 1999 Roadside | 34 ppb |
| Annual hourly Max NO ₂ | 1999 Roadside | 119 ppb |
| Annual Average SO ₂ | 1999 Roadside | 3 ppb |

2.2 Outer London

2.2.1 Bromley Jubilee Country Park (TQ 436 680) Distance from Charing Cross 18 km

This country park is a secondary woodland with remnants of ancient woodland, pasture and cultivated areas covering approximately 25 hectares. It has many small mixed areas of oaks, ash and willow supported by open glades with old field boundaries, hedges and ponds enhancing the open aspect. The western end of the park is dominated by hawthorn with elder, pine and birch. Exceedences of the nitrogen dioxide Objectives are not expected in Bromley, unlike most other London boroughs, with background levels modelled at 15ppb. Additional modelled data on NO₂ by the local authority provides the following information:

| NO ₂ Annual Average Roadside | 16 ppb |
|---|--------|
| NO ₂ Hourly Maximum | 55 ppb |
| SO ₂ Annual Average | 3 ppb |

2.2.2 Enfield Covert Way Nature Reserve (TQ 263 974) Distance from Charing Cross 17 km

This secondary woodland covering approximately 10 hectares, is set within a quiet sheltered suburban location close to Hadley Common. It is rich in mature oak and ash which are quite densely planted in parts, with a small area of grassland in the centre. A return to coppicing within the site has created a more open aspect complementing the denser plantings in this small conservation area, maintaining a good bryophyte flora and a haven for avian populations.

| 1999 NO ₂ Annual Mean Background | 18 ppb |
|---|--------|
| 1999 NO ₂ Hourly Background Max | 90 ppb |
| 1999 NO ₂ Annual Mean Roadside | 24 ppb |
| 1999 NO ₂ Hourly Roadside Max | 93 ppb |
| 1999 SO ₂ Annual Mean | 3 ppb |

2.2.3 Harrow Canons Park, Stanmore (TQ 183 915) Distance from Charing Cross 16 km

This is an 18th Century Estate with some formal features retained, but now managed as open parkland comprising a narrow mixed broadleaf woodland area, open aspect trees, including both ancient and recently planted oaks and a spinney of younger oak, ash, sycamore and elm. Some planted garden areas and ornamental features remain.

| 1999 NO ₂ Annual Mean Suburban | 18 ppb |
|---|--------|
| 1999 NO ₂ Hourly Max Suburban | 86 ppb |
| 1999 SO ₂ Annual Average | 3 ppb |

3. METHODS

3.1 Tree selection

Ash was selected as the major phorophyte as it is widely distributed across London, with oak as the second species. Due to time and budget constraints only six sites were selected and surveys of ash carried out at all sites with oak being examined at Regent's park only. Eight bark pH measurements were taken per tree. Trees were selected to meet the following criteria: unbranched below 200 cm, upright, open aspect, without injury or disease with a minimum girth of 50 cm and a maximum girth of 150 cm. They were selected at a minimum distance of 150 metres from the park entrance in order to reduce roadside influence. The first five trees meeting the selection criteria were surveyed.

3.2 Lichen Sampling Strategy

The approach follows that of Asta et al. (2002). A narrow five-laddered quadrat was attached to the tree at a height of 150-cm using stainless steel pins at each intersection. The quadrat comprises five grid squares, each measuring 10 x 10 cm extending to cover 50 cm in depth. It was positioned at four orientations, North, South, East and West. All lichen species in each quadrat were recorded and given a value of 1, allowing a maximum frequency score of 5 per quadrat and 20 per tree for each species. The Lichen Diversity Value (LDV) for each site is then calculated by adding the frequencies of all species in each quadrat segment and dividing by the number of trees examined (5). In addition, all species below 50 cm and all species above 50 cm up to 200 cm were recorded and any special features noted.

3.3 Bark Acidity Measurements

Bark pH readings at positions 1 and 5 in the quadrat were measured using a flathead electrode (HI 8014 pH meter and BDH Gelplas double junction Flat Tip electrode 309/100/09) according to Looney & James (1988) and Farmer et al. (1990). The bark was moistened with a solution of KCI (0.1 mol) and the reading taken after three minutes, immediately following a second application of KCI.

3.4 Bryophytes, Algae, Fungi and Liverworts

All epiphytes were recorded

3.5 Species identification

Three techniques were employed for identification: Field examination using a x10 hand lens and chemical tests (Orange, 2002) Microscopical examination of thalli and spores Thin Layer Chromatography (Orange, 2002)

The nomenclature follows lists maintained by Brian Coppins (British Lichen Society) (see http://www.argonet.co.uk/users/jmgray/syn.htm)

3.6 Statistical methods

Correlations were calculated using Excel. Species composition and frequency was investigated using multivariate ordination analysis that serves to reduce complex speciessite data to a form that is visually interpretable. While all multivariate analyses were based on the raw data, additional analyses were performed on transformed (double square root; presence-absence) to detect any improved relationships when effects of dominance were reduced. The methods used were:

Non-Metric Multidimensional Scaling (MDS): data from the 10 stations were subjected to analysis using MDS that is currently widely used in the analysis of spatial and temporal change (e.g. Warwick & Clarke, 1991). The recorded observations from the 6 stations were exposed to computation of triangular matrices of similarities between all pairs of samples. The similarity of every pair of sites was computed using the Bray-Curtis index on the raw and transformed data. Clustering was by a hierarchical agglomerative method using group average sorting, and the results are presented as a dendrogram and as a two-dimensional ordination plot.

SIMPER: the MDS clustering program was used to analyze differences between sites. SIMPER (Plymouth Marine Laboratories PRIMER package) enables those species responsible for differences to be identified by examining the contribution of individual species to the similarity measure.

CONPLOT: this program (Plymouth Marine Laboratories PRIMER package) permits environmental variables to be superimposed on MDS plots, giving a visual indication of correlation between clusters and environmental factors (i.e. pH).

4. RESULTS

4.1. Lichens

Species Diversity

A total of 56 different species were recorded. The data are presented in Table 1. (Appendix A)

- Highest diversity was recorded on ash at the two outer London sites of Harrow and Enfield, with 35 species at each site. The third outer London site, Bromley yielded 29 species.
- The three inner London sites all carried 20 species.
- 32 species were recorded on oak in Regents Park.

Fraxinus : 24 species

Arthonia spadicea, Bacidia arceutina, B. laurocerasi, B. naegelii, Caloplaca phlogina, Candelariella reflexa, C. vitellina, Cliostomum griffithii, Dimerella pineti, Diploicia canescens, Hyperphyscia adglutinata, Hypotrachyna revoluta, Lecanora dispersa, L. muralis, Lecanora saligna, Micarea prasina, Phlyctis argena, Physcia aipolia, P. caesia, P. dubia, Punctelia ulophylla, Rinodina subexigua, R. exigua and Strangospora pinicola.

Quercus: 6 species

Lecanora albella, Parmelina tiliacea, P. saxatilis, Physconia grisea, Pleurosticta acetabulum, Rinodina gennarii.

Common to both tree species: 26 species

Amandinea punctata, Bacidia delicata, Candelaria concolor, Evernia prunastri, Flavoparmelia caperata, Flavoparmelia soredians, Hypogymnia physodes, Lecanora carpinea, L. chlarotera, L. conizaeoides, L. expallens, L. symmicta, Lecidella elaeochroma, Lepraria incana, Melanelia subaurifera, Parmelia sulcata, Parmotrema chinense, Phaeophyscia orbicularis, Physcia adscendens, P. tenella, Punctelia subrudecta, Ramalina farinacea, Scoliciosporum chlorococcum, Xanthoria candelaria, X. parientina and X. polycarpa.

Species common to all sites: Amandinea punctata, Bacidia delicata, Parmelia sulcata, Physcia adscendens and P. tenella.

4.2 Lichen Diversity Values (LDV's)

Highest values were recorded at Harrow, followed by Southwark, and Bromley (Appendix A, Table 2):

4.3. Other corticolous flora on Fraxinus excelsior

12 Bryophytes:
Amblystegium serpens, Brachythecium rutabulum, Dicranoweisia cirrata, Dicranum scoparium, Eurhynchium praelongum, Grimmia pulvinata, Hypnum cupressiforme, H.andoi, Orthotrichum affine, O.diaphanum, O.lyellii, Ulota crispa.
2 Parasitic Fungi:
Lachnella alboviolascens, Gloniopsis praelonga
2 Liverworts:
Lophocolea bidentata, Frullania sp.
4 Algae:
Desmococcus viridis, Athelia arachnoidea, Prasiola crispa, Trentepohlia sp.

4.4 Statistical Analysis

4.4.1 Species Numbers

Dendrograms based on species presence on ash at each site (Appendix C, Figure 1) and ash and oak combined demonstrate that similarity percentage contributions lead to a division at the sixty percent level into inner and outer London sites for ash. Flora on the oaks in Regent's Park are more closely aligned to the Outer London sites.

4.4.2. Species Frequency

In all cases quadrat segments (NSWE) cluster together in the 3 principal groups, apart from the north segment quadrat at Tower Hamlets. Bromley trees initially separate at the eighteen percent level (i.e. these are the most distinct) with two further major groups: the first comprising, Southwark, Regents and Tower Hamlets (apart from the north segment) with Enfield, Harrow and the north segment of Tower Hamlets closely aligned (Appendix C, Figure 2).

4.4.3 Species below 50 cm

The dendrogram (Appendix C, Figure 3) suggests a difference between those sites with more dense housing located away from residential areas and those of a more densely populated location where the dog zone would be most influential in creating an eutrophicated substrate in addition to nitrification from anthropogenic emissions. *Xanthoria parientina* and *X. polycarpa* were not recorded in Bromley although they were widespread and frequent at the other sites and are typical species of nitrogen enriched environments. Further details on the origins of the designation 'nitrophyte' and 'acidophyte' can be found in the Section 2: Assessing the role of biological monitoring using lichens to map excessive ammonia (NH₃) deposition in the UK, Appendix 1.

4.5 Girth and pH.

The girth and pH measurements for the sites are given in Table 3 (Appendix A). Ranges on ash were from 3.7 to 5.59 and 2.7 to 5.19 on Oak. The lowest Ash pH was recorded in Bromley. The data presented in Figure 4.5 illustrate that Ash trees at Bromley have a significantly lower bark pH as determined by a Spearman rank correlation coefficient (p = 0.01).

4.6 Vitality

Few signs of poor health were recorded except at Canons Park, Stanmore, where some of the macrolichens were dying. However, the largest thalli of *Parmelia sulcata* and *Flavoparmelia caperata* were measured at this site. Thalli of *Hypogymnia physodes, Evernia prunastri* and *Ramalina farinacea* were generally below normal at less than 0.5 cm. Thalli of *Physcia* species at the Enfield site were numerous, but in the region of 0.5 cm, possibly just colonising the recently coppiced woodland area compared with the very well developed populations at the inner London sites.

5. DISCUSSION

This pilot study investigated the distribution of corticolous lichens at six sites in London. As so few sites have been studied results should therefore be interpreted with care. However there are some interesting results that require further consideration. The species number is very high for such a small sample of only thirty-five trees and the species recorded include many not seen for over fifty years, as well as some never recorded in London before (James et al. 2002). In particular, the macrolichen *Pleurosticta acetabulum* and the crustose *Strangospora pinicola* represent a flora uncommon to urban environments in recent decades.

5.1 Temporal and Spatial Trends

Species numbers have increased significantly over the past fifty years and correlate with falling concentrations of sulphur dioxide (all below the critical level of 8 ppb) and increasing concentrations of nitrogen and other transport emitted pollutants as demonstrated in Figs 1 and 2.

Fig. 1





Laundon (1970) analysed species diversity and distribution in London in the late 1960s. He observed the decline in urban populations to a single species in central London, *Lecanora dispersa*, and a total of nine corticolous species were recorded throughout London (an area defined within a radius of 16 km from Charing Cross).

Significant increases in populations were recorded in 1980 (Rose & Hawksworth 1981) and 1988 (Hawksworth & McManus, 1988) with total corticolous and lignicolous species on mixed tree species across fifty sites recorded as 49, 17 of which were found in Regent's Park. Both these and more recent surveys (Bates et al 1990, 1992, 2001) have referred to the slow reinvasion rates for specific species in London, and noted a change in community structure with the widely distributed species *Hypogymnia physodes* and *Parmelia saxatilis* decreasing and an increase in *Flavoparmelia caperata* and *Physcia aipolia*.

Spatial distribution suggests that inner London ash carry a significantly lower species range than ash in outer London areas (Fig. 3). The use of a single phorophyte for this study has identified a trend not obvious from other recent studies when lichen diversity on mixed tree species and



Fig. 3

other substrates are recorded. The young oaks were in a very sheltered position but

nevertheless suggest that dispersal mechanisms are bringing a rich diversity of species into the centre of London.

The inner London sites carry many of the characteristics of the *Xanthorion* (James, 1974) community and include a high percentage of species favouring high nitrogen environments. The communities recorded on the Oaks resemble more closely those of the *Parmelia caperata –Pertusaria spp.* community described as characteristic of open woodland: *Lecanora chlarotera, Flavoparmelia caperata, Parmotrema chinense, Parmelia saxatilis* and *P. sulcata.*

5.2 Bark pH and Girth

Lichens are very sensitive to bark pH and many species are limited to specific ranges. Using two phorophytes therefore increases the range of potential diversity.

It is interesting to note the lower bark pH of the ash in Bromley where transport emissions and dog zone influences were at their lowest within the study area

The pH and girth of the oaks demonstrate their young age and may account for the large number of species recorded, contrasting sharply with the small number of species on mature oaks in Regent's Park (communicated, Davies L.) and Kensington Gardens (Bates et al 1990, 2001).

5.3 Air Quality

Annual average concentrations of sulphur dioxide across the area are within the recommended Objectives for sensitive vegetation. Nitrogen dioxide concentrations in Inner London range from 50 at roadside locations and 31 at background locations to a low of 14 ppb at outer London sites. Data for nitric oxide are not included in the study but concentrations in London would add significantly to the total NOx concentration as highlighted by Clapp & Jenkin (2001).

6. CONCLUSIONS

The large number of species recorded in this pilot survey demonstrates a significant increase in lichen diversity in London in recent decades.

However, a comparison of species on ash suggests a major difference in species number and composition between the inner and outer London sites. We have identified a trend towards increasing diversity with distance from Charing Cross. The importance of using the same substrate for comparative purposes is highlighted and further enhanced by the diversity on young oaks in Regents Park (James et al. 2002). This study utilises for the first time the proposed EU Quantitative Lichen monitoring methodology. The approach provides a standardised recording technique to assess lichen diversity as an indicator of environmental change.

Both nitric oxide and nitrogen dioxide are toxic to sensitive species, but very few studies have investigated this effect. Concentrations of NO are higher in central London and frequently exceed nitrogen dioxide concentrations.

The high number of 'nitrophytes' suggests that oxidised nitrogen is readily assimilated by many lichen species and is encouraging species considered indicative of eutrophicated areas, particularly on the urban fringe. These results are related to the data collected in other parts of this study.

The disappearance of some species preferring a more acidic substrate was noted during this study.

The survey suggests that species diversity in London is perhaps not as limited by slow dispersal mechanisms as previously considered, although development may be arrested in sensitive species as demonstrated by the frequency of small thalli of several species such as *Evernia prunastri and Ramalina farinacea* and their rarity at other sites The presence of fertile species reproducing by sexual reproduction only (designated F, in Table 1, Appendix A) is interesting and requires further evaluation. An intriguing possibility is that some lichens in London may have existed in forms unrecognisable to lichenologists waiting until conditions are appropriate for development of typical thalli, a phenomenon previously suggested for the lichen *Xanthoria parietina* by Ott (1987). Most lichens identified, however, are able to reproduce asexually using isidia (I) or soredia (S), detachable outgrowths on the thallus containing the photobiont.

7. RECOMMENDATIONS

- The study area should be extended to cover more sites in London to investigate further the association between lichen diversity and oxides of nitrogen identified by this pilot study. In particular the study should include roadside locations where the impact of NO will be highest, contrasting with the background locations selected in this study.
- Species demonstrating a limited tolerance range should be selected for study at a physiological level.
- This study suggests that sensitive species may respond differently to oxidised nitrogen across a range of concentrations and in different forms. These aspects require further investigation particularly in relation to the Objectives for Sensitive Vegetation and Ecosystems. London provides an ideal laboratory to study the effects of oxidised nitrogen. Ambient concentrations of nitrogen dioxide are the highest in the UK (GLA 2001), nitric oxide is particularly high in inner London and ozone and ammonia generally lower than in rural environments.
- The results of this part of the study support the need for a joint project to investigate further the uptake and effect of oxidised and reduced nitrogen on sensitive plant species in relation to current critical levels for nitrogen dioxide and the impact of ammonia.
- The EU draft protocol for lichen biomonitoring used in this report, whilst offering a valuable protocol for quantifying lichen diversity, requires further evaluation concerning assessment of nitrogen emissions.
- The information gained through the above studies should be used as a precursor to developing practical indicator scales and in the development of their regulatory role to complement physicochemical monitoring and modelling studies.

APPENDIX A

Table 1 Species Diversity Summary. N= nitrophytes; A = acidophytes; F = fertile; S = sorediate; I= isidiate

| Summary | Regents park | South- wark | Tower Hamlets | Enfield | Harrow | Bromley | Total | Regents park | Total | N & A | F&S |
|-----------------------------|-----------------|----------------|------------------|---------|--------|---------|-------|-----------------|--------|-------|--------|
| | Fx | Fx | Fx | Fx | Fx | Fx | Fx | | All | | |
| Amandinea punctata | 1 | 1 | 1 | 1 | 1 | 1 | 6 | 1 | 7 | | F |
| Arthonia spadicea | | | | 1 | | | 1 | | 1 | | F |
| Bacidia arceutina | | | 1 | 1 | | | 2 | | 2 | | S |
| Bacidia delicata | 1 | 1 | 1 | 1 | 1 | 1 | 6 | 1 | 7 | | S |
| Bacidia laurocerasi | | | | 1 | | | 1 | | 1 | | F |
| Bacidia naegelii | | | | 1 | | | 1 | | 1 | | F |
| Caloplaca phlogina | | | | | | 1 | 1 | | 1 | | S |
| Candelaria concolor | | | | 1 | | | 1 | 1 | 2 | | S |
| Candelariella reflexa | | 1 | 1 | 1 | 1 | 1 | 5 | | 5 | N | S |
| Candelariella vitellina | 1 | 1 | | 1 | 1 | 1 | 5 | | 5 | N | F |
| Cliostomum griffithii | | | | 1 | | 1 | 2 | | 2 | | F |
| Dimerella pinetii | | | | 1 | | 1 | 2 | | 2 | | F |
| Diploicia canescens | | | | | 1 | | 1 | | 1 | | S |
| Evernia prunastri | | 1 | 1 | 1 | 1 | 1 | 5 | 1 | 6 | Α | S |
| Flavoparmelia caperata | | 1 | | 1 | 1 | 1 | 4 | 1 | 5 | | S |
| Flavoparmelia soredians | | | | | 1 | | 1 | 1 | 2 | | S |
| Hyperphyscia adqlutinata | | | | 1 | 1 | 1 | 3 | | 3 | | S |
| Hypogymnia physodes | | | | | 1 | 1 | 2 | 1 | 3 | А | S |
| Hypotrachyna revoluta | | | | 1 | • | 1 | 2 | | 2 | | F |
| l ecanora albella | | | | | | | 0 | 1 | 1 | | F |
| Lecanora carninea | | | | | 1 | 1 | 2 | 1 | 3 | | F |
| Lecanora chlarotera | | | | 1 | 1 | 1 | 3 | 1 | 1 | | F |
| | 1 | 1 | | 1 | | 1 | 3 | 1 | 4 | Δ | SE |
| Lecanora dispersa | 1 | 1 | 1 | 1 | 1 | 1 | 6 | 1 | 6 | N | 51 |
| | 1 | 1 | 1 | 1 | 1 | 1 | 0 | 1 | 5 | IN | C C |
| | 1 | | | - 1 | - 1 | 1 | 4 | 1 | 1 | NI | 5 |
| | 1 | | | 4 | | | 1 | | 1 | IN | |
| | | | | 1 | 1 | | 1 | 1 | 2 | | г Г |
| | | | 4 | 1 | 1 | 1 | 2 | 1 | 3 F | | F |
| | | | 1 | 1 | 1 | 1 | 4 | 1 | 5 | • | F |
| Leprana mcana/iop | | 4 | 1 | 1 | 1 | 1 | 4 | 1 | 5 | A | 5 |
| Melanella subaurifera | | 1 | 1 | 1 | 1 | 1 | 5 | | 6 | | 3 |
| Micarea prasina | | | 1 | | | | 1 | 4 | 1 | • | SF |
| Parmelia saxalliis | 4 | 4 | 4 | 4 | 4 | 4 | 0 | 1 | 1 | A | 1 |
| Parmella suicata | 1 | 1 | 1 | 1 | 1 | 1 | 6 | 1 | 1 | | 5 |
| Parmelina tiliacea | | | | | | | 0 | 1 | 1 | | 1 |
| Parmotrema chinese | | | | 1 | 1 | 1 | 3 | 1 | 4 | | 8 |
| Phaeophyscia orbicularis | 1 | 1 | 1 | 1 | 1 | | 5 | 1 | 6 | N | 5 |
| Phlyctis argena | | | | 1 | | | 1 | | 1 | | S |
| Physcia adscendens | 1 | 1 | 1 | 1 | 1 | | 4 | 1 | 5 | N | 5 |
| Physcia alpolia | 1 | 1 | | | 1 | | 3 | | 3 | | F |
| Physcia dubia | | | | | 1 | 1 | 2 | | 2 | N | S |
| Physcia sp.* | 1 | 1 | 1 | 1 | 1 | 1 | 6 | | 7 | | S |
| Physcia tenella | 1 | 1 | 1 | 1 | 1 | 1 | 6 | 1 | 7 | N | S |
| Physconia grisea | | | | | | | 0 | 1 | 1 | | S |
| Physica caesia | | | | | 1 | | 1 | | 1 | N | S |
| Pleurosticta acetabulum | | | | | | | 0 | 1 | 1 | | F |
| Punctelia ulophylla | | | | | 1 | | 1 | | 1 | | S |
| Punctelia subrudecta | 1 | 1 | | | 1 | 1 | 4 | 1 | 5 | | S |
| Ramalina farinacea | 1 | | | 1 | 1 | 1 | 4 | 1 | 5 | | S |
| Rinodina gennarii | | | | | | | 0 | 1 | 1 | N | F |
| Rinodina subexigua | 1 | | | | | | 1 | | 1 | | F |
| Rinodina exigua | | | 1 | | 1 | | 2 | | 2 | | F |
| Scoliciosporum chlorococcum | 1 | 1 | 1 | 1 | 1 | | 5 | 1 | 6 | | SF |
| Strangospora pinicola | | | | 1 | 1 | 1 | 3 | | 3 | | F |
| Xanthoria candelaria | 1 | 1 | 1 | 1 | 1 | 1 | 5 | 1 | 6 | N | S |
| Xanthoria parietina | 1 | 1 | 1 | 1 | 1 | | 5 | 1 | 6 | N | F |
| Xanthoria polycarpa | 1 | 1 | 1 | 1 | 1 | | 5 | 1 | 6 | N | F |
| | 20 | 20 | 20 | 35 | 35 | 28 | | 32 | | | |

| | North | South | East | West | Totals |
|---------------|-------|-------|------|------|--------|
| Westminster | 16.8 | 9.6 | 6.3 | 11.0 | 43.6 |
| Southwark | 12.8 | 16.0 | 11.8 | 9.4 | 50.0 |
| Tower Hamlets | 8.6 | 11.2 | 8.4 | 10.8 | 39.0 |
| Harrow | 11.4 | 19.0 | 15.6 | 14.2 | 60.2 |
| Bromley | 4.5 | 22.8 | 8.8 | 12.8 | 48.9 |
| Enfield | 10.2 | 15.0 | 8.8 | 11.0 | 45.0 |

Table 2. Frequency Values: Lichen Diversity Values (Asta, 2002)

The Lichen Diversity Value is calculated by taking the sum of the frequency of each species as recorded in the total quadrat for the relevant orientation and dividing by the number of trees per site. The totals are then summed to give a final diversity score (lichen diversity value 'LDV') per site

| | | R | egen | ts | |] | Towe | r Ha | mlet | S | Southwark | | | | | Enfield | | | | Harrow | | | | | Bromley | | | | | |
|------------|-------|--------|--------|------|------|-------|-------|------|-------|-------|------------|-------|--------|--------|-------|---------|-------|-------|-------|--------|--------|------|------|-------|---------|------|-----|-----|-----|-----|
| | R1 | R2 | R3 | R4 | R5 | T1 | T2 | T3 | T4 | T5 | S 1 | S2 | S3 | S4 | S5 | E1 | E2 | E3 | E4 | E5 | H1 | H2 | H3 | H4 | H5 | B1 | B2 | B3 | B4 | B5 |
| Ash | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Gir | 96 | 77 | 107 | 156 | 82 | 65 | 58 | 65 | 52 | 71 | 76 | 79 | 90 | 84 | 92 | 116 | 127 | 70 | 58 | 70 | 50 | 68 | 82 | 91 | 46 | 99 | 108 | 91 | 76 | 133 |
| pН | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| S 1 | 4.5 | 5 | 5.1 | 4.4 | 4.4 | 5.5 | 5.6 | 4.8 | 4.7 | 5.4 | 4.3 | 4.5 | 5 | 5.3 | 4.7 | 4.9 | 5.6 | 4.9 | 5.1 | 4.9 | 5 | 5 | 5 | 5.3 | 5 | 4.3 | 4.4 | 4.5 | 4.2 | 4.5 |
| S5 | 5.2 | 4.9 | 5.3 | 5.2 | 4.6 | 5.5 | 5.2 | 5.1 | 4.9 | 5.4 | 4.9 | 5 | 5 | 5 | 4.8 | 4.5 | 5.4 | 5.1 | 5 | 5.2 | 5 | 4.9 | 5 | 5.1 | 4.9 | 4.9 | 4.7 | 4.6 | 3.9 | 4.7 |
| E1 | 4.7 | 4.7 | 5.2 | 4.7 | 4.7 | 5.6 | 4.9 | 5.2 | 5 | 5.1 | 4.9 | 4.5 | 5.1 | 5.3 | 5 | 5 | 5.8 | 5.1 | 5.1 | 5.1 | 5.1 | 5.1 | 5 | 5.3 | 4.8 | 3.6 | 4.3 | 4.1 | 3.8 | 4.9 |
| E5 | 5 | 5 | 5.2 | 4.2 | 4.9 | 5.5 | 5.2 | 5.3 | 5.1 | 5.3 | 5 | 4.9 | 5.4 | 5.2 | 5.1 | 5.4 | 5.5 | 5.1 | 5 | 5 | 4.9 | 5.2 | 5.2 | 5.2 | 5 | 4.1 | 4.4 | 4.2 | 3.9 | 4.9 |
| N1 | 4.8 | 4.9 | 5.2 | 4.3 | 4.9 | 5.8 | 5.4 | 5.1 | 5.1 | 4.9 | 4.6 | 4.5 | 5.2 | 5.6 | 5.3 | 5.5 | 5.5 | 5.2 | 5.1 | 5.5 | 4.9 | 5 | 5 | 5.2 | 4.2 | 3.6 | 4.2 | 3.7 | 4.2 | 3.9 |
| N5 | 5 | 4.9 | 5.1 | 4.2 | 4.7 | 6 | 5.4 | 5.1 | 5.1 | 5.1 | 4.8 | 4.8 | 4.8 | 5.4 | 5.3 | 5.8 | 5.6 | 5.2 | 5.1 | 5.6 | 4.9 | 5.1 | 4.9 | 5 | 4 | 4.7 | 4.2 | 3.7 | 3.6 | 4.3 |
| W1 | 4.8 | 5 | 4.9 | 4.4 | 4 | 5.5 | 5.5 | 5.1 | 5.3 | 5.5 | 4.4 | 4.7 | 5.1 | 4.7 | 5.2 | 5.3 | 5.6 | 5.3 | 5.3 | 4.9 | 5.1 | 5.2 | 5.4 | 5.9 | 4.7 | 3.9 | 4.3 | 4 | 3.8 | 4.6 |
| W5 | 4.9 | 5.2 | 5.3 | 4.3 | 4.1 | 5.8 | 5.4 | 5.1 | 5.4 | 5.4 | 4.9 | 5.5 | 4.8 | 4.8 | 5.3 | 5.7 | 5.7 | 5.3 | 5.5 | 4.8 | 5 | 5.2 | 5.4 | 5.3 | 4.8 | 4.5 | 4.6 | 4.3 | 4 | 4.4 |
| | | 0 | ak | • | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Gir | 51 | 51 | 62 | 98 | 53 | | | | | | | | | | | | | | | | | | | | | | | | | |
| pН | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| S 1 | 4.2 | 4.1 | 4.2 | 3.2 | 5 | | | | | | | | | | | | | | | | | | | | | | | | | |
| S5 | 4.2 | 4.3 | 4.2 | 3.8 | 4.9 | | | | | | | | | | | | | | | | | | | | | | | | | |
| E1 | 4 | 4.4 | 4.7 | 3.3 | 4.8 | | | | | | | | | | | | | | | | | | | | | | | | | |
| E5 | 4 | 4.4 | 4.7 | 3.9 | 5.2 | | | | | | | | | | | | | | | | | | | | | | | | | |
| N1 | 4.3 | 4.6 | 4.4 | 3.3 | 4.7 | | | | | | | | | | | | | | | | | | | | | | | | | |
| N5 | 4 | 4.2 | 4.3 | 2.7 | 5.1 | | | | | | | | | | | | | | | | | | | | | | | | | |
| W1 | 4.5 | 4.3 | 4.5 | 3.4 | 4.7 | | | | | | | | | | | | | | | | | | | | | | | | | |
| W5 | 4.5 | 4.1 | 4.5 | 3 | 4.7 | | | | | | | | | | | | | | | | | | | | | | | | | |
| pНı | eadi | ngs - | S1 i | ndic | ates | posit | ion 1 | in q | uadr | at (1 | 0x10 |) cm |) at a | heig | tht o | f 150 |) cm(| top o | of qu | adra | t), S5 | indi | cate | s pos | sition | 5 at | | | | 1 |
| base | of q | uadr | at at | a he | ight | of 60 | cm | (top | of qu | ladra | at fac | ing S | South | ı (the | en Ea | ast, N | Jorth | , Ŵe | est) | | | | | 1 | | | | | | |
| (top | of q | uadra | at fac | cing | Sout | h | | | | | | | | , | | | | | | | | | | | | | | | | |
| (the | n Eas | st, No | orth, | West | t) | | | | | | | | | | | | | | | | | | | | | | | l | | |

Table 3. Girth and pH measurement

APPENDIX B

Figure 1.1 Modelled Concentrations of Nitrogen Dioxide in London (ppb) 1999 (1997 weather data) Source: Greater London Authority



APPENDIX C

Figure 1. Species number on Ash by site (PRIMER): A = Outer London Sites. B = Inner London



Figure 2. Ash Frequency Data (PRIMER)



Figure 3. Similarity below 50 cm (Dog Zone) (PRIMER)



Figure 4. Ash pH ranges (PRIMER)



Ash

8. REFERENCES

- Air Quality Framework Directive (Council Directive 96/62/EC) See <u>http://www.europa.eu.int/comm/environment/air/ambient.htm</u>.
- AEA Technology/NETCEN (1998) Annual Report.
- Asta, J., Erhardt, W., Ferretii, M., Fornasier, F., Kirschbaum, U., Nimis, P.L., Pirintsos, S., Purvis, O.W., Scheidegger, C., Van Haluwyn, C. & Wirth, V. (2002). Mapping Lichen Diversity as an Indicator of Environmental Quality. In Nimis, P.L., Scheidegger, C. & Wolsely, P.A. (Eds.) Monitoring with Lichens. Kluwer, Dordrect: 273-279.
- Bates, J.W., Bell, J.N.B. & Farmer, A.M. (1990). Epiphyte recolonisation of oaks along a gradient of air pollution in South-east England, 1997-1990. *Environmental Pollution* **68:** 81-99.
- Bates, J.W. & Farmer, M.A. (Eds.) (1992). *Bryophytes and Lichens in a Changing Environment*. Clarendon Press, Oxford.
- Bates, J.W., Bell, J.N.B. & Massara, A.C. (2001). Loss of *Lecanora conizaeoides* and other fluctuations of epiphytes on oak in S.E. England over 21 years with declining SO₂ concentrations. *Atmospheric Environment* **35**: 557-568.
- Bromley Local Authority (2000). Review and Assessment of Air Quality in Bromley. Environmental Health Department, Bromley Council.
- Clapp, L.J. & Jenkin, M.E. (2001) Analysis of the relationship between ambient levels of O₃, NO₂ and NO as a function of NOx in the UK. *Atmospheric Environment* **35**: 6391-6405.
- DETR (2000) The Air Quality Strategy for England, Scotland, Wales and Northern Ireland. Working Together for Clean Air. Department of the Environment, Transport and the Regions, in partnership with the Scottish Executive, The National Assembly for Wales, and the Department of the Environment for Northern Ireland. Available on-line at

http://www.environment.detr.gov.uk/airquality/index.htm

- Farmer, A.M., Bates, J.W. & Bell, J.N.B (1990). A comparison of methods for the measurement of bark pH. *Lichenologist* **22**: 191-197.
- Hawksworth D.L., & McManus P. (1989). Lichen recolonisation in London under conditions of rapidly falling sulphur dioxide levels and the concept of zone skipping. *Bot. J. Linn.Soc.* **100**: 99-109.
- James, P.W., Purvis, O.W. & Davies, L. (2002). Epiphytic Lichens in London. *Bulletin of the British Lichen Society.* **90:** 1-3.
- Laundon, J.R. (1970). London's Lichens. The London Naturalist: 49: 2-69.
- Looney, J.H. & James, P.W. (1988). Effects on lichens. In *Acid Rain and Britain's Natural Ecosystems* (M.A. Ashmore, J.N.B. Bell & C. Garretty, eds): 13-25. London: Imperial College Centre for Environmental Technology
- Mayor's Draft Air Quality Strategy (2001). Greater London Authority (GLA), Romney House, Marsham Street, London SW1.

- Orange, A., James, P.W., White, F.J. (2002). Microchemical methods for the identification of Lichens, British Lichen Society
- Ott, S (1987) Reproductive strategies in lichens. *In:* E. Peveling (ed.): *Progress and Problems in Lichenology in the Eighties*. Bibliotheca Lichenologica No. 25. J. Cramer, Berlin-Stuttgart, pp. 81-93.
- PRIMER, Plymouth Routine in Marine Environmental Research. Non-metric multidimensional scaling (Bray Curtis 1957).
- Rose, C.I. & Hawksworth, D.L. (1981). Lichen recolonisation in Cleaner Air. *Nature* **289**: 289-292.
- Rose, F. & James, P.W. (1974). Regional Studies on the British Lichen Flora 1. The corticolous and lignicolous species of the New Forest, Hampshire. *Lichenologist* **6**: 1-72
- SEIPH (1999) Annual Report. Air Quality in London, SEIPH-ERG, King's College, London.
- Southwark Local Authority, Diffusion Tube Survey 1997.
- Tower Hamlets (2002) Nitrogen dioxide modelled contour map.
- Warwick, R.M. & Clarke, K.R. (1991) A comparison of some methods for analysing changes in benthic community structure. J. mar. biol. Ass. U.K., **71:** 225-244.
- World Health Organisation Air Quality Guidelines for Europe (2000). Published by WHO, Regional Office for Europe, Scherfigsvej 8, DK-21-00 Copenhagen, Denmark

PART 2: INVESTIGATING THE IMPACT OF NOx ON TRANSPLANTS

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1. Introduction

Lichen transplants are widely used to assess the impact of pollution in areas where native thalli are rare or absent. Numerous chemical analytical studies have been carried out including monitoring lead from roads, heavy metals from incinerators and large-scale studies investigating spatial and temporal patterns in metal deposition, as well as assessing the impact of pollutants on various physiological parameters (Garty 2001). The national German guideline VDI 3799 relies on a visual assessment of damage to lichen thalli to evaluate phytotoxic effects. As lichens are notoriously sensitive to environmental change, including local climatic factors, recent guidelines emphasise a need to transplant lichens from sites having ecologically similar environments to those of the transplant sites (Mikhailova 2001).

Monitoring studies in London have identified a decrease in SO₂ tolerant species (e.g. Bates *et al.* 2001). At Burnham Beeches, Buckinghamshire, 40 km to the west of London, the appearance of species such as SO₂ sensitive species such as *Xanthoria* and the shrubby lichens (*Evernia prunastri* and *Ramalina*) were noted on oak trees (Purvis *et al.* 2001a,b). Photographic monitoring recorded an impact on lichen growth (*Parmelia sulcata*) during episodic high pollutant emissions, coupled with unusual climatic conditions which suggests that traffic emissions may be responsible for impacts on lichen floras (Purvis *et al.* 2001a,b). This was supported by chemical analysis of samples demonstrating the accumulation of Zn, N, and other signature elements of traffic pollution. These studies suggested a combination of nitrogen and/or particles may play a role in influencing lichen growth. *P. sulcata* is one of the most widely employed lichen species in Europe to monitor spatial and temporal patterns in metal deposition.

The Natural History Museum's Wildlife Garden situated next to the junction of Cromwell Road, is infamous for its high traffic density and pollution

concentrations (Fig. 1). The DEFRA air pollution monitoring station, part of the automatic pollution recording network is





housed in its western corner at the Queen's Gate intersection (Fig. 2). Kensington and

Chelsea Environmental Health department has also established weekly dust monitoring guages. Regular exceedances of objectives for human health for NO_2 (21 ppb annual mean) and particles led the Royal Borough of Kensington and Chelsea to declare the borough as an Air Quality Management Area. Imperial College in association with Newcastle, Bradford and Manchester Metropolitan Universities and the Centre for Ecology and Hydrology (CEH) at Bangor is currently conducting a survey to monitor the effect of urban air pollution on vegetation creating an ideal opportunity for the current pilot project.

2. Objectives

To establish lichen transplants in the NHM wildlife garden in the vicinity of existing $N0_x$, 0_3 , VOC monitoring gauges in order to evaluate the impact of urban air pollution on epiphytic twig communities over a 3-month period:

- To assess the abundance and vitality of 3 lichen groups: fruticose and SO₂ sensitive (*Ramalina/Evernia*) and the foliose *Parmelia sulcata* (SO₂ tolerant) and *Xanthoria parietina* (N tolerant).
- To sample lichen transplants (*Parmelia sulcata* and *Xanthoria parietina*) along the pollution gradient and retain material for future chemical analysis
- To compare results with physicochemical data and make recommendations for future work.

3. Materials and Methods

Lichens on twigs were selected as transplant materials since:-

- Twigs have been successfully used as transplants both to detect temporal and spatial patterns in metal deposition (e.g. Garty 2001).
- Lichen communities on twigs are particularly sensitive to air pollution (Wolseley 1999)
- The majority of thalli on twigs are young, in active growth and therefore less susceptible to senescence through natural causes
- Twigs established on a pole creates a near natural structure where stem flow effects are minimised

Sample collection was carried out during the first week of December 2001 at Nettlecombe, Somerset. Samples were collected from the outer and lower reachable branches of a single isolated Sycamore tree (Acer pseudoplatanus). This species has a neutral/alkaline bark pH and has the closest affinity with the London Plane (Platanus acerifolio) surrounding the Wildlife Garden. Approximately 70 twigs aged between 2-8 years and ca. 40-80cm in length were collected and 50 of these mounted on bamboo radial arms supported on five 7 cm diameter pvc pipes. Other more mature twigs (ca.15) aged between 4-18 years from a different sycamore tree were placed on plastic nets at 3 sites at about 5 m from the road along the southern and western boundary of the wildlife garden (Fig. 2). The winter period was selected for our experiment owing to shading / barrier effect by trees and increased the microclimatic variation during the Summer period. 20 sticky pads were sited adjacent to the trees at each site according to a method developed by Dr Ben Williamson at the Natural History Museum: The garden is situated below the Cromwell and Queens Gate roads:

Garden



Cromwell Road

| Monitoring Station | Distance from Cromwell road | Aspect |
|--------------------|-----------------------------|---|
| Site 1 | 20 m | S-facing, exposed |
| Site 2 | 30m | SW, exposed also to Queen's Gate |
| | | road |
| Site 3 | 60m | Sheltered by other vegetation |
| Site 4 | 90m | exposed to Queen's Gate road to the west but sheltered on other sides |
| Site 5 | 120m | S- and W- facing on the north side lies NHM building and Springhouse |

Table 1. Transplant site locality details

Changes in the composition of species groups was assessed at the start and end of the transplant period according to the following indices:

| Frequency Index | 1 = a single individual | 2 = 1-5 thalli | 3 = 6-10 thalli | 4=11-20 thalli | 5 > 20 thalli |
|--------------------|--------------------------------------|------------------------|--------------------------|------------------------|---------------|
| Vitality index | 1 = healthy (no signs of | 2 = slight signs of | 3 = moderate signs of | 4 = necrotic (100%) | |
| | bleaching or necrotic lesions) | necrosis (<30%) | necrosis (>30-50%) | | |
| Class size | < 1 cm = small 's' | 1-3 cm = medium 'm' | > 3 cm large 'l' | | |

Table 2. Indices to quantify lichen transplant frequency, vitality and class size

Chlorophyll fluorescence provides a rapid method to determine lichen vitality in the field (Jensen 2002, Jensen & Kricke 2002). Chlorophyll fluorescence was carried out on 12 & 14 January 2001 at the NHM wildlife Garden by Randolph Kricke using a Walz PAM-210 (teaching PAM) (see http://www.walz.com/pamzhsp.htp. Measurements were repeated using a Hansatech Plant Efficiency Analyser after a 3 month exposure period in the Wildlife Garden (15 March) by Sarah Honour and Annalisa Massara and at the transplant site at Somerset (17 March) by Feliciano Cirimele. Chlorophyll fluorescence was carried out on a minimum of 6 *Parmelia sulcata* thalli at each station with reference to the parameter Fv/Fm on moistened and previously dark adapted thalli. Dark adaptation was carried out in January using black plastic and in March with dark adaptation clips. Measurements were taken on young margins in all cases.

Sample collection for chemical analysis

Samples of *P. sulcata* and *Xanthoria parietina* were collected for analysis. Owing to the major loss of *P. sulcata* towards the centre of the wildlife garden, samples of *Evernia* and *Ramalina* were also taken with a view to performing an inter-species calibration.

Digital photography

Photographs were taken at monthly intervals of 6 thalli of *Xanthoria parietina* and *Parmelia sulcata* at each site using a NIKON Coolpix 995 camera. Images were taken at a fixed distance 7 cm using manual focus.

4. Results

Lichens at the control site in Nettlecombe remained healthy throughout the exposure period, both based on photographic recording and measurements of chlorophyll fluorescence. Growth was recorded in both *Parmelia sulcata* and *Xanthoria parietina* demonstrating that the lichens were successfully transplanted at Nettlecombe.

Changes in lichen frequency

In NHM wildlife garden, an assessment of the frequency of different size classes in the 3 species groups revealed different trends across the transect (Fig. 3). Least changes were observed in the most exposed sites (1 and 2) adjacent to road-sites (Figs 3A & B) where fewer losses were recorded than 3 and 4 (Figs 3C & D) (sheltered) and 5 (Fig. 3E) (exposed and furthest from roadsides). Maximum losses of all species groups (> occurred in large thalli) at site 3 (Fig. 3C), the most sheltered site towards the centre of the garden. Virtually all





Parmelia sulcata disappeared from this site. The increase in numbers of small thalli recorded in sites 4 and 5 reflect the disintegration of larger thalli leaving marginal remnants.

Changes in vitality

Minor signs of thallus bleaching or the development of a characteristic pinkish or brownish discoloration was observed at any site. Least changes in lichen vitality were recorded at the exposed sites 1 and 2 in all species groups as determined by a visual assessment (Fig. 4) and digital photography. Indeed, images of lichens from Site 2 at the level of Queen's Gate road showed the least changes.



Fig. 4. Average Vitality Index

This contrasted with sheltered sites where major losses occurred. Exposed site 5 furthest from the road suffered the most significant loss of thalli of *Parmelia sulcata*, particularly towards the inner thallus centre and the site appeared to be deteriorating in a similar fashion to sites 3 and 4. *Xanthoria parietina* showed the opposite behaviour to *Parmelia sulcata* with younger thalli suffering the major loss, especially from sites 3 and 4.



Chlorophyll fluorescence measurements for *Parmelia sulcata* transplants, NHM Wildlife Garden and thalli at Somerset (transplants and native thalli)

Fig. 5

Chlorophyll fluorescence

Considerable variation in chlorophyll fluorescence as measured by Fv/Fm was noted (*Fig. 5*). Most striking is the observation that the Somerset specimens have much lower values (March 2002) than those transplanted to NHM wildlife garden – including measurements made in January 2002 one month after transplantation. The standard error is also considerably larger for March measurements. Measurements confirm that all lichens are physiologically active and alive.

Digital photography

Photographic recording confirms that growth has occurred in transplants at the Somerset control site. Little change was noted in thalli at exposed sites at the NHM wildlife garden (1 and 2). Major loss was noted at sites 3 and 4. Older thalli of *Parmelia sulcata* after 6 weeks of exposure showed a major loss of inner parts at sites 3, 4 and 5. In contrast, larger *Xanthoria* thalli tend to loose the outer part at the same sites (*Appendix E*).

5. Discussion

A combination of a visual assessment of abundance, vitality, digital photography and chlorophyll fluorescence confirms that the healthiest lichens are those transplanted at sites 1 and 2 adjacent to the Cromwell and Queen's Gate roads. Maximum N0, N0₂, N0x and VOC air concentrations were recorded at these sites suggesting that NOx concentrations which exceed the health standards for human health are not having a

major impact on lichen health over the short term (3 months). This contrasts with previous studies where major impacts were recorded on lichens exposed to pollution. Data on the effects of NOx on lichen vitality in the field are lacking although Gilbert (1968) recorded major reductions in chlorophyll content (84%) and respiration (80%) in transplants of the lichen *Ramalina farinacea* at mean SO₂ concentrations of 264 μ g/m³ (Gilbert 1968) during the survey. The loss of lichens in the sheltered sites might be due to a number of reasons quite apart from pollution including:

- Microclimate: Increased shelter will result in prolonged hydration (and hence metabolic activity) during which various pollutants may be in solution. Unfavourable microclimatic factors would result in degeneration in the absence of pollution. December remained dry and relatively warm. Under these conditions lichens would remain physiologically less active. However, rainfall increased during January and February when lichens would be metabolically active and most sensitive to damage from pollution.
- Other biological factors including Long-tailed tits and other birds for use in nesting materials, grazing by invertebrates.

Chlorophyll fluorescence

Caution is advisable over the interpretation of fluorescence data owing to the different equipment used with potentially different sensitivities, the different operators and methods used to dark adapt thalli. If there is a relationship there appears to be a tendency towards higher levels within the wildlife garden. However, levels above 0.75 are rare in lichens (Jensen 1992) and this upward trend contradicts previous studies mainly examining impacts of SO₂ resulting in a decrease in Fv/Fm (see Jensen 2001). SO₂ is well known to result in chlorophyll pigment degradation in lichen photobionts. However, Von Arb & Brunold (1990) identify higher chlorophyll contents in lichens exposed to traffic pollution. There is no significant difference between Fv/Fm values obtained for NHM wildlife garden site 2 (exposed to traffic) and transplant site 1 in Somerset.





Fig. 7

6. Conclusion

The pilot project confirms that epiphytic lichens on twigs can be successfully introduced into London to study pollution effects even where NO and NO₂ levels are high. The absence of a major direct impact on the lichens at road-sites adjacent to Cromwell Rd and Queen's Gate suggests that NOx is not having a major impact on lichen growth over this time period. This suggests that N may not, at least at these concentrations and meteorological conditions, have an effect on the vitality of *P. sulcata* although, a longer exposure period would certainly be required to fully test this hypothesis. The possible effect of VOC's and particulates also needs to be considered. It is interesting to note that the shrubby lichens *Evernia* and *Ramalina* remained largely unaffected in this region. There was clearly a varied response amongst the different species and thalli of different ages. We suggest that in addition to pollution, microclimatic factors may have played a role in determining the vitality and loss of particular species. A larger study over a longer time period is required.

7. Recommendations

- It is now important to carry out chemical analysis of the lichen transplants and bark samples we collected during the experiment for N and a range of emission relevant elements. It will be interesting to analyse the lead levels following reductions in the use of leaded petrol. Characterisation of the chemical and physical form of particulates trapped on the sticky pads by SEM and comparison with metal accumulation by lichens at the same sites.
- Further transplant experiments should be established across a wider NOx gradient in London to complement biodiversity studies in areas where insufficient native thalli are available for analysis to determine possible critical levels under different meteorological conditions.
- Further research needs to be carried out to test and develop chlorophyll fluorescence as an indicator of NOx air pollution both in relation to transplants and natural thalli.
- Detailed growth measurements (e.g. Hill 2002) can be carried out on lichens provided sufficient thalli are available for measurements and growth coefficients calculated. In addition to transplants, this can usefully be extended to the study of natural populations as little is understood of the dynamics of natural populations in London.

Appendix A

Estimated Air Pollution Data for the Wildlife Garden, 7th November 2001 to 15th March 2002

Nitrogen Oxides

Concentrations of nitric oxide (NO) and nitrogen dioxide (NO₂) at the 5 sites in the Wildlife Garden were estimated by calculating the relationship between measured site concentrations and the NOx levels recorded by the DEFRA monitoring station at the corner of the garden. This relationship was then applied to data from the monitoring site for the time period when the lichen were exposed.

The relationship between NOx concentrations at the sites in the Wildlife Garden and those measured by the DETR monitoring site were calculated from existing data (See Appendix 1). The NO₂ relationship was calculated from 4 sets of measurements made with Palmes type diffusion tubes and 3 sets of measurements made with Ogawa NO/NOx passive samplers. The NO relationship was calculated from 2 sets of measurements made with Ogawa NO/NOx passive samplers. The relationship from the 3rd set of measurements made with these samplers were not used in the calculations since they were taken during a period of high NO concentrations and gave a different relationship to the other measurements. The NO levels recorded at the DEFRA monitoring site during the experiment were similar to those used to estimate the relationship.

| | DEFRA | | | | | |
|----------------------------|---------|--------|--------|--------|--------|--------|
| | station | Site 1 | Site 2 | Site 3 | Site 4 | Site 5 |
| NO % | 100 | 34 | 32 | 18 | 20 | 20 |
| NO conc.(ppb) | 75 | 25 | 24 | 13 | 15 | 15 |
| | | | | | | |
| NO ₂ % | 100 | 67 | 66 | 57 | 52 | 57 |
| NO ₂ conc.(ppb) | 36 | 24 | 24 | 21 | 19 | 21 |
| | | | | | | |
| NOx conc.(ppb) | 111 | 50 | 48 | 34 | 34 | 36 |

Figure 1: Estimated NO and NO₂ concentrations at the 5 sites in the Wildlife garden, 07/11/01 - 15/03/02

Appendix B

Measurements used to estimate the NO and NO₂ relationship for the 5 Sites in the Wildlife Garden

| | Sampling | | Si | Site 1 | | te 2 | Si | te 3 | Si | te 4 | Site 5 | |
|------------|--------------|-----------|----------------|------------|----------------|------------|----------------|------------|----------------|------------|----------------|------------|
| | Dates | DEFR A | Conc. (ppb) | % DEFRA | Conc. (ppb) | % DETRA | Conc. (ppb) | % DETRA | Conc. (ppb) | % DETRA | Conc. (ppb) | % DETRA |
| Nitrogen | Dioxide | | (11), | | | | | | | | U I J | |
| Palmes | 12-26/07/00 | 44 | 27 | 61 | 26 | 59 | 23 | 52 | 20 | 45 | 36 | |
| type | 26/07-23/08 | 42 | 33 | 79 | 32 | 76 | 32 | 76 | 27 | 64 | 20 | 48 |
| diffusion | 23/08-22/09 | 44 | 32 | 73 | 34 | 77 | 22 | 50 | 18 | 41 | | |
| tubes | 22/09-20/10 | 47 | 32 | 68 | 26 | 55 | 26 | 55 | 24 | 51 | 29 | 62 |
| Ogawa | 18/10-01/11 | 37 | 24 | 65 | 24 | 65 | 18 | 49 | 19 | 51 | 18 | 49 |
| passive | 01/11-16/11 | 39 | 27 | 69 | 27 | 69 | 26 | 67 | 24 | 62 | 27 | 69 |
| samplers | 19/11-3/12 | 41 | 23 | 56 | 24 | 59 | 20 | 49 | 20 | 49 | 23 | 56 |
| | Relationship | | | 67 | | 66 | | 57 | | 52 | | 57 |
| Nitric Oxi | de | | | | | | | | | | | |
| Ogawa | 18/10-01/11 | 82 | 28 | 34 | 27 | 33 | 14 | 17 | 13 | 16 | 16 | 20 |
| passive | 01/11-16/11 | 103 | 45 | 44 | 39 | 38 | 41 | 40 | 30 | 29 | 33 | 32 |
| samplers | 19/11-3/12 | 83 | 28 | 34 | 26 | 31 | 16 | 19 | 20 | 24 | 16 | 19 |
| | Relationship | | | 34 | | 32 | | 18 | | 20 | | 20 |

SOMERSET TRANSPLANT SITE

January 2002





Parmelia sulcata



Xanthoria parietina











NHM WLDLIFE GARDEN TRANSPLANT SITE 2 (Xanthoria parietina)



Appendix F





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8. References

- Bates, J.W., Bell, J.N.B., & Massara, A.C. (2001) Loss of *Lecanora conizaeoides* and other fluctuations of epiphytes on oak in S.E. England over 21 years with declining SO₂ concentrations. *Atmospheric Environment* **35**: 2557-2568.
- Garty, J. (2001) Biomonitoring atmospheric heavy metals with lichens: theory and application. *Critical Reviews in Plant Sciences* **20(4):** 309-371.
- Gilbert, O.L. (1968) Biological indicators of air pollution. PhD thesis. University of Newcastle UponTyne.
- Hill, D.J. (2002) Measurement of lichen growth. In *Protocols in Lichenology: Culturing, Biochemistry, Ecophysiology and Use in Biomonitoring.* Kranner, I., Beckett, R.P & Varma, A.K., eds.: 255-278, Springer, Berlin.
- Jensen, M. (2002) Measurement of chlorophyll fluoresence in lichens. In Protocols in Lichenology: Culturing, Biochemistry, Ecophysiology and Use in Biomonitoring. Kranner, I., Beckett, R.P & Varma, A.K., eds.: 136-151, Springer, Berlin.
- Jensen, M. & Kricke, R. (2002) Chlorophyll fluorescence measurements in the field assessment of the vitality of large numbers of lichen thalli, In Nimis, P.L., Scheidegger, C. & Wolseley, P.A., eds, *Monitoring with lichens-Monitoring Lichens* Kluwer. Academic Publishers 327-332.
- Mikhailova, I. (2001) Transplanted lichens for bioaccumulation studies. In Nimis, P.L., Scheidegger, C. & Wolseley, P.A., eds, *Monitoring with lichens Monitoring Lichens*. Kluwer, Dordrecht: 301-304.
- Purvis, O.W., Bamber, R.N., Chimonides, J., Erotokritou, L., Jones, G., Jeffries, T. & Din, V. (2001a) *Burnham Beeches Lichen Monitoring Phase 2.* Year 3. Report for Corporation of London.
- Purvis, O.W., Chimonides, J., Din, V.K., Erotokritou, L., Jeffries, T., Jones, G.C., Louwoff, S., Read, H. & Spiro, B. (2002) Which factors are responsible for the changing lichen floras of London? *Detecting Environmental Change. Science and Society*, 17-20 July, London, UK. Pp 24-25.
- Purvis, O.W., Chimonides, J, Din, V., Erotokritou, L., Jeffries, T. Jones, G.C., Louwhoff, S., Read, H., Spiro, B. Which factors are responsible for the changing lichen floras of London? *Science and the Total Environment* [in press]
- Purvis, O.W., Erotokritou, L., Wolseley, P.A., Williamson, B. & Read, H. (2001) A photographic quadrat recording method employing digital analysis of lichens as an indicator of environmental change. In Nimis, P.L., Scheidegger, C. & Wolseley, P.A., eds, Monitoring with lichens – Monitoring Lichens. Kluwer, Dordrecht: 337-341.
- VonArb, C. & Brunold, C. (1990) Lichen physiology and air pollution. 1. Physiological response of in situ *Parmelia sulcata* among air pollution zones within Biel, Switzerland. *Canadian Journal of Botany.* **68:** 35-42.

- Wolseley, P.A. (2002). Using lichens on twigs to assess changes in ambient atmospheric conditions. - In: Nimis, P.L. et al. (eds.): Monitoring with Lichens – Monitoring Lichens. NATO Science Series, IV, vol. 7. Kluwer, Dordrecht, pp. 291-294.
- Wolseley, P.A. & Pryor, K.V. (1999). The potential of epiphytic twig communities on *Quercus petraea* in a welsh woodland site (Tycanol) for evaluating environmental changes. *Lichenologist* **31**: 41-61.

SECTION 2

Assessing the Role of Biological Monitoring Using Lichens to Map Excessive Ammonia (NH₃) Deposition in the UK

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D. Table of epiphytic lichens on *Quercus* trunks at North Wyke, Devon
E. Table of species recorded on *Quercus* twigs.
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1. BACKGROUND

Research on isolated oak trees in the vicinity of intensive livestock units in the Netherlands has shown a strong correlation between the composition of lichen communities and atmospheric ammonia levels (van Herk 1999, 2002). The recording technique identified species that were obligate nitrophytes (nitrogen-loving species) or acidophytes (SO₂ tolerant species) enabling the construction of indices of nitrophyte 'NIW' (Nitrofiele Indicatie Waarde) and acidophytes 'AIW' (Acidofiele Indicatie Waarde).

In Britain, the Ammonia monitoring network at the Centre for Ecology and Hydrology, Edinburgh (http://www.nbu.ac.uk/cara/UKNAMN/uknamn.htm), monitored ammonia levels at 95 sites. Recent research in the vicinity of intensive poultry units in Britain has shown high levels of ammonia deposited locally along a deposition gradient of up to c. 300m from source (Fowler *et al.*, 1998). The residence time of ammonia in the atmosphere is rather brief so that the effects on vegetation are often localised around the source (NEGTAP 2001).

Although lichens have long been shown to be indicators of atmospheric acidification (Nimis *et al.*, 2002), recent observations on epiphytic lichen communities in rural areas in the Netherlands have shown a widespread change in lichen communities with an increase in nitrophytes and a corresponding decrease in acidophytes (van Herk, 1999). Whereas there is little variation in the climate of the Netherlands, there is a marked climatic gradient across Britain ranging from drier conditions in the east to wetter oceanic conditions in the west which have a pronounced influence on the species composition of lichen communities. The continental method developed by van Herk was applied during the present survey in the contrasting climatic regions of Norfolk and Devon, in the vicinity of ammonia monitoring stations in order to test the potential application of the system in Britain.

2. OBJECTIVES

- To collect data from 2 sites in Britain with a history of intensive livestock management where NH_3 emissions have been monitored over a period of time.
- To test the results against scales developed in the Netherlands.
- To evaluate the method in order to develop a biomonitoring procedure in Britain.

3. METHOD

3.1. Site selection

Sites were selected using aerial photographs, taking into account: the availability of ammonia monitoring stations; the distribution of lichens in Britain including both continental and oceanic components; the presence of exposed oak trees on acid soils; and the presence of a site of conservation importance in the vicinity.

A site with a continental climate was selected in the Thetford region, Norfolk, where extensive ammonia monitoring has been taking place in the vicinity of intensive poultry

and pig rearing units and in unimproved grassland utilised by the MOD. A site with an oceanic climate at North Wyke in Devon was chosen where ammonia is recorded in the vicinity of intensive cattle rearing sheds at IGER (Institute of Grassland and Environmental Research).

3.2. Station selection

Stations were selected according to availability of *Quercus* trees and prevailing wind direction. The distance to source of pollution was estimated from ordnance survey maps. Managed woodland and woodland edge trees were avoided where possible, as lichen communities varied considerably in these sites.

Stations were selected in Norfolk to coincide with existing ammonia monitoring stations. At North Wyke stations were selected within zones of up to 1000m radius of the farm units, as the ammonia station is located south and upwind of the cattle sheds and slurry tank.

Given the restrictions of time and the availability of trees, 3-5 upright standard *Quercus* trees >50cm girth were selected in exposed well-lit situations. Leaning trees were avoided or those with low epicormic growth or ivy on the trunk. Trees on the edge of managed woodland or intensively used roads were also avoided. Stations and tree position were marked on a map. Bark samples were collected for pH determination.

3.3. Lichen recording.

Corticolous species on each tree trunk up to 2m were listed and assigned a frequency value according to van Herk; 1 thallus on the trunk, 2 or more thalli, and species with >10cm² present. Where identification was not possible in the field specimens were collected for identification in the laboratory.

The data was subsequently converted for each station according to van Herk.

- a) Only one thallus present
- b) 2 or more thalli on one tree
- c) Present on 30-50% of trees, < 10cm²/tree
- d) Present on 30-50% of trees, > 10cm²/tree
- e) Present on 50-100% of trees, <10cm²/tree
- f) Present on 50-100% of trees, >10cm²/tree
- g) NIW and AIW species were distinguished and scored according to van Herk and the mean AIW and NIW scores for each station were calculated (Appendix 1).

3.4. Lab techniques

Bark pH.

3 samples of bark were collected from each tree, placed in paper packets and dried. pH was measured using a GELPLAS flathead electrode after moistening bark with 25% KCl for c. 5 minutes (Bates *et al.* 1990).

Lichen species were identified using light microscopy and thin layer chromatography

3.5. Data analysis

Ammonia deposition was supplied as monthly averages for all stations in the Thetford area by the Breckland Council for Environmental Health and by CEH Edinburgh ammonia

for the period February to October 2001 and as monthly averages for North Wyke by CEH Edinburgh between May 1997 and September 2001. Where present, mean ammonia values for lichen sampling stations in the Thetford area were plotted against NIW and AIW indices. As ammonia deposition was only recorded at one station at North Wyke, the distance from the farm units and slurry tank was plotted against NIW and AIW indices as a surrogate measure for ammonia deposition.

4. RESULTS

Site 1: Thetford area, Norfolk

In Norfolk 12 stations were sampled in the Thetford area coinciding where possible with ammonia monitoring stations (Fig1, table 1).

| | | | NH ₃ deposition | | | | Lichen div | ersity | | |
|------|----------------------------|--------|----------------------------|------|-------|-----|------------|--------|---------------|----------------|
| Stat | Station location | NH_3 | High | Mean | Low | no. | Trunk | Twig | Distance from | Direction from |
| | | no. | | | | T's | | | source | source |
| 1 | Norfolk Wildlife Reserve 1 | 11 | 2.64 | 2.88 | 0.8 | 5 | 7.4 | 5 | 500 | SW |
| 2 | Abrey field | c.13 | 3.13 | 2.9 | 1.86 | 3 | 10 | 2.7 | 300 | All round |
| 3 | West Tofts | 20 | 2.7 | 1.9 | 0.99 | 4 | 8.5 | 6.6 | 8000 | W |
| 4 | Thorpe Gt heath | c.9 | 2.95 | 2.2 | 1.32 | 3 | 7.7 | 7.7 | 2600 | W |
| 6 | Military camp | 10 | 3.91 | 2.8 | 1.44 | 3 | 7 | 3 | 2-400 | SW & NE |
| 7 | Hockham | 25 | 124.7 | 43 | 27.27 | 4 | 7 | 4 | 2-300 | SW &NE |
| 8 | Peddars way adj. Poultry | 16 | 16.46 | 9.2 | 5.63 | 3 | 5.3 | 0.5 | 50 | E |
| | units | | | | | | | | | |
| 9 | Peddars way adj. pigs | c.22 | 9.81 | 4.54 | 1.4 | 3 | 10.3 | 2 | 50 | All round |
| 10a | Norfolk Wildlife Reserve 2 | 12 | 4.45 | 1.6 | 1.92 | 2 | 7.2 | 3.5 | 800 | SW |
| 10b | Norfolk Wildlife Reserve 3 | 15 | 2.91 | 1.9 | 1.12 | 2 | 7.2 | 5.5 | 1300 | N & E |
| 11 | Stanford area | c.19 | 2.03 | 1.4 | 0.86 | 3 | 10 | 6.1 | 7000 | NW |
| 12 | Santon Downham FC | 1 | 3.32 | 2.05 | 1.2 | 3 | 7.3 | 8 | 8000 | W |

Table 1. Stations in the Thetford area.

Fig. 1. Map of Thetford area showing NH_3 recording stations (see Appendix 1 for map of site with lichen stations).



Lichen diversity

A total of 45 lichen species were recorded in the Thetford site with mean lichen numbers varying from 4.7 to 10.7 species per station (Appendix 1). The lowest diversity coincided with proximity to intensive poultry units where ammonia levels were highest (32-124 µg/m³ in October 2001) (fig. 2.). Overall the highest diversity of lichens occurred on oaks on MOD land west of the poultry units in unimproved grassland where stocking rates were low (12,000 sheep on 27,000 acres (pers. comm.)). Lichen diversity on twigs showed a better correlation with ammonia deposition than lichen diversity on trunks (figs. 2 & 3.). Contrary to expectations, diversity was often highest on younger tree trunks and lowest on trees of great girth and age in all stations (fig. 4.), even on MOD land where there has been no intensive agriculture for more than 50 years. This suggests previous damage to older trunks probably from a long period of sustained acidification. Within the MOD area foliose species of the Parmelietum were present and often abundant in the canopy, but on the trunks they were absent or restricted to the lowest parts of the tree trunk.

Diversity was also high in arable areas, although the community was dominated by nitrophyte species of the Xanthorion.

Distribution of nitrophyte and acidophyte species.

Only 9 NIW and 4 AIW species were recorded in the Thetford site in Norfolk from 20 possible nitrophyte and 20 acidophyte species characterised by van Herk. Mean ammonia deposition in all stations adjacent to lichen sampling was plotted against indices of AIW and NIW for these stations (Figs. 5 & 6). The NIW index showed a strong correlation with ammonia levels (Fig. 5), and the AIW showed little correlation with ammonia levels (Fig. 6). However in stations adjacent to the highest ammonia deposition lichen diversity was 0 -1



















on both trunks and twigs suggesting that there are critical loads of ammonia at which nitrophytes are absent.

Bark pH was also strongly affected in those areas where NH₃ levels exceeded $10\mu g/m^3$ (Fig. 8). The stations in unimproved grassland in MOD or Forestry Commission sites with mean bark pH's between 3 and 4.50 coincided with ammonia deposition below 3 (except site 12 where lichen sampling was in unimproved pasture upriver from the ammonia recording site which was by a horse-riding centre without oak trees). In contrast mean bark pH's >5 were associated with stations where ammonia levels were above 10 $\mu g/m^3$ (fig. 7). The local ammonia deposition gradient was confirmed by bark pH measurements at the Norfolk Wildlife's Reserve (NWT), where bark pH was up to 6.2 in the vicinity of the poultry units and fell to c. 4 in the vicinity of Langmere 400m away. The directional nature of deposition was also indicated by variation in pH around the trunk on one young *Quercus* in the vicinity of the poultry unit, where a pH of 6.2 on the N. side of the trunk contrasted with a pH of 3.9 on the S side.





Site 2. North Wyke IGER, Devon.

In Devon 7 sampling stations were studied at North Wyke (Fig. 8, Table 2.) and 77 lichen species recorded (see Appendix 2.).

| | | • | | • | | | |
|---------|-------|----------|----------|--------|------|------|-----|
| | | | | Lichen | | | |
| | | | | | Sity | | |
| Station | No. | Distance | Directio | trunk | twig | NIW | AIW |
| no | trees | from | n from | | _ | | |
| | | source | source | | | | |
| 2 | 5 | -20 | E | 18.2 | 5 | 0.8 | 1 |
| 1 | 5 | 30 | W | 18 | 4.4 | 0.4 | 1.4 |
| 3 | 4 | -300 | E | 15.7 | 9.3 | 0 | 0 |
| 4 | 4 | 300 | W | 20.7 | 11.5 | 1.25 | 3 |
| 5 | 3 | 750 | NW | 18.6 | 12.3 | 0 | 5 |
| 6 | 2 | 150 | SW | 16.5 | | 0 | 1 |
| 7 | 4 | 700 | W | 18 | 11.2 | 0 | 3.5 |

Table 2. Stations sampled at North Wyke, Devon

Distribution of stations sampled at North Wyke shown in Appendix 2

Lichen diversity

Lichen diversity per tree was high in all stations, averaging 17.6 on trees east of the cattle sheds and slurry tank to 20.7 on trees in pasture west of the sheds. Ammonia data was only available for one site to the west of the farm unit and levels were low throughout the year the highest being $3.54 \ \mu g/m^3$ during August and September in each year. As the main concentration of ammonia is from the overwintering cattle sheds and slurry tank east of the ammonia monitoring station, distance from source was used as a measure of likely ammonia distribution along an east to west transect. The prevailing wind is from the southwest (figs. 8 & 9).

Mean lichen diversity on *Quercus* trunks per station plotted against distance from source on an east west axis shows little positive correlation (Fig. 8). Lichen diversity of twigs was considerably lower in the vicinity of the cattle sheds and slurry tank and was positively correlated with distance from source in both eastern and western directions (Fig. 9).

This site also supports 10 lichen species that are indicators of long ecological continuity (NIEC species) as defined by Rose (1992), including the highly pollution-sensitive *Lobaria pulmonaria**, *Teloschistes flavicans** and *Usnea ceratina*. These species occur in well-lit sites with high relative humidity and absence of either acidification or eutrophication (Table 3.). Other indicators of long ecological continuity include *Cresponea premnea* and *Lecanographa lyncea*, both species characteristic of





dry bark. These species were fragmented and in poor condition restricted to the densely

shaded drier E. side of the trunk. These ancient woodland indicators are now almost all restricted to a single tree on the boundary of the settlement (S6) and as such are relicts of a former wood pasture regime.

| No. of trees at station | 5 | 4 | 5 | 4 | 4 | 3 | 1 |
|-------------------------|----|----|----|----|----|----|----|
| | S2 | S3 | S1 | S4 | S7 | S5 | S6 |
| Arthonia vinosa | | | | | | 2 | |
| Dimerella lutea | | | 2 | | | | |
| Cresponea lyncea | | | | | | | 2 |
| Lecanographa premnea | | | | | | | 1 |
| Lecanora jamesii | 2 | | | 2 | 1 | | |
| Lobaria pulmonaria | | | | | | | 2 |
| Punctelia reddenda | | | 6 | | 1 | | |
| Phaeographis dendritica | 1 | 1 | | | | | |
| Teloschistes flavicans | | | | | | | 2 |
| Usnea ceratina | | | | 3 | 2 | 3 | |
| No of NIEC species | 2 | 1 | 2 | 2 | 3 | 2 | 4 |
| | | | | | | | |

Table 3. Distribution of NIEC species in stations at North Wyke stations in order of distance from source

NIW and AIW distribution.

12 AIW and 6 NIW species were recorded at North Wyke from a total of 20 AIW 20 NIW species as defined by van Herk. However species of the Xanthorion in the NIW were rare or absent on trunks but present at low to high levels on twigs at all sites, except in one sampling station on an acid wetland reserve where acidophytic species were dominant and the nitrophyte species of the Xanthorion absent. There is little correlation between distance from the farm units and NIW species (Fig. 11) while AIW species show a strong correlation – (R²=0.9) with distance from source suggesting that the farm unit has an effect on these species.

Bark pH.

Bark pH for all trees sampled showed little variation between trees at each station at North Wyke (Fig. 12). The lowest bark pH occurred at station 5 on the edge of a *Molinia* moor. If bark pH is plotted against distance from source, there is a trend towards higher pH in the vicinity of the farm unit and a rapid fall off within 400m in both an easterly and westerly direction (Fig. 13). This suggests that the farm unit is responsible for the observed changes in bark pH.











5. DISCUSSION

Variation between sites.

Both sampling sites were on acid substrata, one on sand (Thetford) and the other on heavy riverine clays and loams (North Wyke), and both sites contained mature oak trees well distributed across the site. The sites are climatically very different. Rainfall in East Anglia averages 55 mm per year with considerable dry periods whereas in Devon the average is c. 176 mm per year with high relative humidity in all months. This allows a wide variety of oceanic lichen species to flourish and lichen cover and diversity tends to be higher in this region.

Previous acidification effects are conspicuous in the Thetford region on the ancient oaks where lichen diversity was very low even in areas of unimproved wood pasture in the MOD area where there has been long ecological continuity. At North Wyke, where there is no evidence of former acidification, large old *Quercus* tree trunks support a highly diverse lichen community even in the vicinity of the farmstead including many oceanic species, pollution sensitive species and indicators of long ecological continuity (Table 1). In this site there was little impact of ammonia on long established natural lichen communities. Although both Thetford and North Wyke are areas with a tradition of wood pasture since Norman times with potentially long ecological continuity, the absence of former acidification and the higher rainfall at North Wyke must contribute to the survival of pollution sensitive species in this site.

Changes in temperature regimes may also contribute to changes in the distribution of lichen species. Many nitrophytes are predominantly warm temperate species and are benefiting from the trend of increasing mean annual temperatures associated with global warming (van Herk *et al.*, 2002).

Nitrophyte and acidophyte species.

Only 9 out of a possible 20 NIW species, and 8 out of 20 possible AIW species listed in van Herk were recorded in the surveys. This is partly due to the bias towards continental species in the Netherlands, and that the UK flora is more oceanic even in the eastern regions. Nitrophytes that are abundant in the Netherlands are absent or rare in Britain and others that may be considered as nitrophytes are excluded (see James et al. 1976). Alternative species considered to be nitrophytes in Britain that are present at the sites visited are indicated by a* in the table of species in the Appendices. Further testing in order to derive an appropriate species list for both nitrophytes and acidophytes is essential in regions where atmospheric conditions are monitored. In the Netherlands several new species have been described that are now appearing in Britain.

Ammonia deposition and lichens.

Ammonia concentrations at stations in the area around Thetford were positively correlated with NIW indices and distance from source and showed little correlation between AIW indices and distance from source.

There was no correlation of NIW species at North Wyke on trunks with distance from the farm unit as very few of the NIW species of the Xanthorion were present in the closed communities on the oak trunks. However, NIW species were present on twigs, indicating

that these species were colonising recently established lichen communities. At North Wyke ammonia levels were low, but the recording site was situated on the western side of the farm unit and would have only received higher levels during the distribution of slurry. The negative correlation of AIW species with distance from the farm unit, indicates that these acidophyte species are affected by the farm unit. The corresponding increase in bark pH in the vicinity of the farm unit correlates with the loss of acidophytic species characteristic of undisturbed lichen communities in this region. Bark pH correlated with lichen communities recorded at both Thetford and North Wyke and to the ammonia levels (Thetford) or distance from source (North Wyke). In areas where there is little environmental change, bark pH's are relatively stable on the tree and at the site. Whereas in areas where changes are occurring as in the Thetford region, there are wide variations in bark pH even on one tree suggesting that there is a directional element in the pollutant causing an increase in bark pH. Increased bark pH is associated with an increase in ammonia deposition. Preliminary results on 3 trees at site 1 & 10 in the NWT reserve show a rapid change in bark pH suggesting that there is an environmental gradient on this site which agrees with Fowler et al. (1998) in falling off at c. 700m from the source. This is not supported by the ammonia deposition tubes which record highest levels of ammonia as $4.2\mu g/m^3$ in a recording station in the vicinity of station 10a.

The positive correlation between ammonia deposition and NIW species at Thetford indicates a similarity between lowland sites in the east of Britain and the Netherlands. It also suggests that there are critically high levels of ammonia in this region where lichens including NIW species are absent (Figs 3 & 5).

Lichen diversity on trunks and twigs

Preliminary observations from a basic inventory of lichen diversity on twigs at both sites suggest that primary colonisers of the newer bark surfaces are the most sensitive to changes in atmospheric conditions. In contrast, trunk species may reflect longer-term environmental conditions of ecological continuity (as at North Wyke) and former acidification through SO₂ air pollution (Thetford). Although there was very little correlation between lichen diversity on trunks and proximity to sources of ammonia at North Wyke and Thetford, there was a strong correlation between lichen diversity on twigs and ammonia sources at both sites (figs. 3 & 9). Nitrophyte species were present in this community. At North Wyke nitrophyte Xanthorion species were conspicuously absent from the trunks but present on the twigs in the vicinity of the farmstead.

6. CONCLUSIONS

This pilot survey using techniques and indicator species defined as nitrophytes and acidophytes during research in the Netherlands (van Herk, 2002) suggests that epiphytic lichen communities on oak are sensitive to increased levels of atmospheric ammonia in Britain.

However many NIW and AIW species recorded for the Netherlands were not found in sites sampled in either Norfolk or in Devon, suggesting that further work is needed to define appropriate nitrophyte and acidophyte indicator species in Britain. The research should define indicator species in relation to the wide climatic gradient across Britain, and in relation to other widespread nitrogen sources such as from fertilisers, car exhausts and fossil fuels, whose effect on lichen assemblages is poorly understood. Lichen species identified as possible candidates for these lists are indicated by a * in Appendices 3 & 4. As with the research in the Netherlands it is to be expected that species new to science or to Britain may be identified during surveys.

The results of a rapid survey of lichens on twigs made during the pilot study is particularly interesting and suggests that there is a strong correlation between species colonising the new bark surface and distance from ammonia sources. Further research is needed together with NH₃ monitoring to test the capacity for twig communities to respond to rapid fluctuations in NH₃ concentrations over short periods of time, compared with communities and NIW/AIW indicators on trunks and twigs. The existence of a sampling method (Wolseley & Pryor, 1999; Wolseley, 2002) will allow a quantitative assessment of change in lichen communities to be tested against ammonia levels and other parameters.

7. RECOMMENDATIONS

- Define nitrophyte and acidophyte species appropriate for the UK based on multivariate analysis.
- Extend quantitative sampling to include twigs on oak trees across a gradient of NH₃ deposition.
- Devise a practical recording method using lichens to detect changes in atmospheric nitrogen appropriate for Britain
- Include recording of bark pH together with ammonia and other nitrogen products monitoring across Britain, with particular reference to areas that were formerly acidified.
- Undertake further research on the effects of agricultural management on lichen communities including research stations where agricultural methods and eutrophication factors are being researched. The high lichen diversity at North Wyke makes this an ideal situation in which to extend ammonia monitoring and establish lichen monitoring.

APPENDIX A

Maps of lichen stations at Thetford and North Wyke.



IGER, North Wyke

APPENDIX B

Species identified by van Herk as nitrophytes and acidophytes (1999, 2002). Species highlighted were found during the survey.

APPENDIX C.

Table of species found on *Quercus* trunks in vicinity of Thetford with van Herk frequency values for each species at each station. Acidophytes in blue, nitrophytes in yellow. *Additional species to be considered as nitrophytes in the UK.

| Order of stations by NH ₃ dep. | | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 |
|---|----------|---------|-----------|------------|----------|-----------|---------|-----------|------------|------------|-----------|-----------|------------|
| Mean diversity per trunk | Stations | 7 S7 | 5.3 S8 | 10.3 S9 | 10 S2 | 7.4 S1 | 7 S6 | 7.7 S4 | 6.1 S12 | 7.5 S10 | 8.5 S3 | 10 S11 | 7.3 S12 |
| Species on trunk | | | | | | | | | ÷ | | | | |
| *Amandinea punctata | | 2 | | 5 | 6 | 1 | 6 | | | 3 | 3 | 3 | |
| Arthonia spadicea | | | | | | | | | 1 | | | | 1 |
| Arthonia sp | | | | | | 1 | | | | | | | |
| Athelia sp | | | | | | 1 | | | | | | | |
| *Bacidia delicata | | 1 | | | | 2 | | 2 | | 2 | | 2 | |
| Bryophytes | | 6 | | 1 | 6 | - | | - | 6 | 6 | 5 | 5 | 6 |
| Candelariella reflexa | | 1 | | - | - | | | | - | - | - | _ | - |
| Candelariella vitellina | | | | | 3 | | | | | | | | |
| *Chrvsothrix candelaris | | | | | | | | | | | 2 | | |
| Cladonia sp. | | | | | | | | | 2 | | | | 2 |
| Cliostomum griffithii | | | 2 | 3 | 1 | | 6 | | 3 | 2 | | 3 | 3 |
| Desmococcus | | | 6 | 6 | 6 | 6 | 6 | 6 | Ū | - | | 6 | · · |
| Dimerella nineti | | | Ū | Ū | Ū. | Ū | U U | · · | 1 | | | Ū | 1 |
| Dinloicia canescens | | 3 | | | 3 | | 2 | | | | 3 | 5 | |
| Evernia prunastri | | 2 | | | 0 | 1 | 6 | 6 | | 3 | 2 | Ũ | |
| *Hyperphyscia adqlutinata | | - | | | 1 | | Ŭ | Ŭ | | Ŭ | _ | | |
| Hypogymnia physodes | | | | | | | | 2 | | | | | |
| cf Lecanora compallens | | 3 | | 5 | 5 | 2 | | | 6 | 6 | 6 | | 6 |
| Lecanora dispersa | | 2 | | 5 | | | | | | | | | |
| *Lecanora expallens | | 1 | 3 | | 2 | | 2 | | | 3 | | 3 | |
| *Lecidella elaeochroma | | 2 | | 1 | | | | | | | | | |
| Lepraria incana | | | 5 | 2 | | 6 | | 6 | 6 | 6 | 6 | | 6 |
| Melanelia subaurifera | | | 1 | | 1 | 3 | 2 | 3 | 2 | 3 | | | 2 |
| Micarea prasina | | | | | | | | | | 2 | | 1 | |
| Haematomma caesium | | | | | 2 | | | | | | | | |
| Opegrapha atra | | | | 1 | | | | | | | | | |
| Flavoparmelia caperata | | | | | | | | | 2 | 2 | 3 | 5 | 2 |
| Hypotrachyna revoluta | | | | | | | | | | | | 2 | |
| Parmelia sulcata | | | | | 1 | | | 6 | | 3 | 2 | 2 | |
| Pertusaria amara | | | | | | | | | | | 2 | | |
| Pertusaria hymenea | | | | | | | | | | | | 2 | |
| Phaeophyscia orbicularis | | | | | | 2 | | | | | | | |
| Phlyctis argena | | | 2 | | | | | | 2 | | 2 | | 2 |
| Physcia adscendens | | 2 | 1 | 5 | | | | | | | | | |
| Physcia tenella | | | 2 | 5 | 6 | 6 | 5 | 3 | | 2 | 3 | | |
| *Physconia grisea | | | | | 5 | | | | | | | | |
| Punctelia borreri | | | | | | | | | | | | 1 | |
| Punctelia subrudecta | | | | | | 3 | 3 | 2 | 2 | | | 3 | 2 |
| Punctelia ulophylla | | | | | | | | | | | | | |
| Pyrrhospora quernea | | | | | | | | | | | | 3 | |
| Ramalina farinacea | | | | | | | 3 | | | 3 | 1 | 1 | |
| Ramalina fastigiata | | | | | | | | 3 | | | | | |
| Schismatomma decolorans | | | | 3 | | | | | | | | 3 | |
| *Scoliciosporum chlorococcum | | 3 | | | | 6 | | | | 5 | | | |
| Xanthoria candelaria | | 3 | 2 | 1 | | 3 | | 2 | | 3 | | 3 | |

| Xanthoria parietina | 6 | 2 | 6 | 6 | 3 | 3 | 2 | | 2 | |
|---------------------|---|---|---|---|---|---|---|---|---|---|
| Xanthoria polycarpa | 3 | 2 | 3 | 5 | 2 | 1 | | 2 | | 2 |
| Total 48 species | | | | | | | | | | |

APPENDIX D

Table of species found on *Quercus* trunks at North Wyke, Devon with van Herk frequency values for each species at each station. Acidophytes in blue, nitrophytes in yellow. *Additional species to be considered as nitrophytes in the UK.

| Stations sampled | S1 | S2 | S3 | S4 | S5 | S6 | S7 |
|-----------------------------|----|------|----|------|----|----|------|
| Av. Species no. per station | 18 | 17.6 | 18 | 20.7 | 18 | 18 | 16.5 |
| | | | | | | | |
| Acrocordia conoidea | | 3 | | | | | |
| Acrocordia gemmata | | | | | | | 2 |
| *Amandinea punctata | 5 | 4 | 3 | 3 | | | |
| Arthonia impolita | | | | | 3 | | |
| Arthonia radiata | 2 | | | | | | 2 |
| Arthonia vinosa | | | | | 2 | | |
| *Bacidia delicata | | | 2 | 3 | | | |
| Bacidia rubella | 1 | | | | | | |
| Candelariella reflexa | | 1 | | | | | |
| Candelariella vitellina | | | | 2 | | | |
| *Chrysothrix candelaris | | 5 | 2 | 3 | 3 | | 4 |
| Cladonia coniocreae | | | | | 1 | | 3 |
| Cladonia pyxidata | | | | | 2 | | |
| Cladonia ramulosa | | | | | 1 | | |
| Cladonia sp. | 2 | | | | 3 | 1 | |
| Cliostomum griffithii | 2 | | 3 | 5 | 3 | | 2 |
| Dimerella lutea | 2 | | | | | | |
| Dimerella pineti | | | 2 | | | | |
| *Diploicia canescens | | 4 | | | | | |
| Enterographa crassa | 1 | 4 | | | | 2 | |
| Evernia prunastri | | 3 | | 3 | | | 3 |
| Fuscidea lightfootii | | | | 2 | | | |
| Hypogymnia physodes | | 1 | | | 2 | | |
| Hypotrachyna revoluta | 6 | 5 | 4 | | | 2 | 3 |
| Cresponea lyncea | | | | | | 2 | |
| Lecanographa premnea | | | | | | 2 | |
| Lecanora chlarotera | 5 | 3 | 4 | 5 | 3 | 2 | 3 |
| Lecanora dispersa | | 1 | | | | | |
| *Lecanora expallens | 6 | 6 | 6 | 6 | 6 | 2 | 6 |
| Lecanora jamesii | | 2 | | 2 | | | 1 |
| *Lecidella elaeochroma | 5 | | 5 | 5 | | 1 | 2 |
| Lepraria sp | 2 | | 4 | 3 | 3 | | |
| Lobaria pulmonaria | | | | | | 2 | |
| Melanelia fuliginosa | | | | | | | 2 |
| Melanelia subaurifera | 5 | 3 | | 3 | | | 3 |
| Normandina pulchella | | | | | | 2 | |
| Ochrolechia subviridis | 5 | 2 | 3 | 3 | | 2 | 2 |
| Ochrolechia turneri | | | 1 | | | | |
| Opegrapha atra | 5 | 6 | 1 | 1 | | 2 | |
| Opegrapha herbarum | | 3 | | | | | |
| Opegrapha multipuncta | | | | | 1 | | |
| Opegrapha vulgata | | | | | | | |
| Flavoparmelia caperata | 6 | 4 | 1 | 5 | 3 | | 6 |
| Parmelia saxatilis | 1 | 2 | | 3 | | | 3 |
| Parmelia sulcata | 2 | 3 | 3 | 3 | 2 | | 2 |

| Parmotrema chinense | 6 | 4 | 4 | 6 | 6 | 2 | 6 |
|-----------------------------------|---|---|---|---|---|---|---|
| Pertusaria albescens v. corallina | 4 | 3 | 6 | | 6 | 2 | 6 |
| Pertusaria albescens | | | 2 | 6 | 2 | | 1 |
| Pertusaria amara | 3 | 3 | 4 | 3 | 3 | | 6 |
| Pertusaria hemispherica | | | | | 2 | | |
| Pertusaria hymenea | 6 | 6 | 6 | 6 | 6 | 2 | 5 |
| Pertusaria pertusa | | | 3 | 5 | 4 | | 6 |
| Phaeographis dendritica | | 1 | 1 | | | | |
| Phaeophyscia orbicularis | | 1 | | | | | |
| Phlyctis argena | | 3 | | 5 | 4 | | 3 |
| *Hyperphyscia adglutinata | | 3 | | 1 | | | |
| *Physcia pulverea | | 3 | | | | | |
| Physcia tenella | | | | 2 | | | |
| Placynthiella icmalea | 1 | | | | | | |
| Punctelia borreri | | | | 1 | | | |
| Punctelia reddenda | 5 | | | | | | 1 |
| Punctelia subrudecta | | 3 | 3 | | | 2 | |
| Punctelia ulophylla | | | 3 | | | | |
| Pyrrhospora quernea | 5 | 5 | 1 | 3 | 6 | | 6 |
| Ramalina calicaris | | | | | | | 1 |
| Ramalina canariensis | 2 | | | | | | |
| Ramalina farinacea | 2 | 1 | 4 | 3 | | | 4 |
| Ramalina fastigiata | 2 | 1 | 3 | 3 | | | 2 |
| Rinodina roboris | | 1 | | | | | |
| Schismatomma cretaceum | | 3 | | | | | |
| Schismatomma decolorans | 6 | 6 | 4 | 5 | 6 | | 6 |
| Teloschistes flavicans | | | | | | 2 | |
| Usnea ceratina | | | | 3 | 3 | | 2 |
| Usnea cornuta | 3 | | | | 2 | | 2 |
| Usnea flammea | | | | 2 | 3 | | |
| Usnea rubicunda | | | | 1 | 1 | | |
| Xanthoria parietina | 2 | 2 | | 1 | | | |
| Total diversity = 78 | | | | | | | |
| Total AIW = 12 | 3 | 3 | 0 | 5 | 9 | 1 | 5 |
| Total NIW = 6 | 1 | 4 | 0 | 3 | 0 | 0 | 0 |

APPENDIX E

Table of species recorded on Quercus twigs Acidophytes in blue, nitrophytes in yellow.

North Wyke

Acrocordia conoidea Amandinea punctata Arthonia punctiformis Arthonia radiata Arthopyrenia fallax Arthopyrenia punctiformis Evernia prunastri Fuscidea lightfootii Graphina anguina Graphis scripta Hyperphyscia adglutinata Hypogymnia tubulosa Hypotrachyna revoluta Lecanora albella Lecanora chlarotera Lecanora expallens Lecanora intumescens Lecanora persimilis Lecanora symmicta Lecidella elaeochroma Lecidella elaeochroma f. soralifera Melanelia subaurifera Micarea prasina Opegrapha atra Opegrapha herbarum Flavoparmelia caperata Parmelia sulcata Pertusaria amara Phaeographis dendritica Physcia aipolea Physcia tenella Punctelia borreri Punctelia subrudecta Ramalina canariensis Ramalina farinacea Ramalina fastigiata Scoliciosporum chlorococcum Usnea subfloridana Xanthoria parietina Xanthoria polymorpha

Thetford

Amandinea punctata Arthonia punctiformis Arthonia sp Arthopyrenia sp. Athelia sp Bacidia delicata Candelariella vitellina Chrysothrix candelaris Cliostomum griffithii Desmococcus Diploicia canescens Evernia prunastri Hyperphyscia adglutinata Hypogymnia physodes Hypogymnia tubulosa Lecanora carpinea Lecanora chlarotera cf. Lecanora compallens Lecanora expallens Lecanora dispersa grp Lecanora expallens Lecidea symmicta Lecidella elaeochroma Lepraria incana Melanelia subaurifera Mycoporum quercus Flavoparmelia caperata Parmelia sulcata Pertusaria amara Phaeophyscia orbicularis Physcia adscendens Physcia aipolea Physcia tenella Phlyctis argena Physconia grisea Punctelia borreri Punctelia subrudecta Punctelia ulophylla Ramalina farinacea Scoliciosporum chlorococcum Xanthoria candelaria Xanthoria parietina Xanthoria polycarpa

REFERENCES

- Farmer, A.M., Bates, J.W. & Bell, J.N.B. (1990) A comparison of methods for the measurement of bark pH. *Lichenologist* **22**: 191-197
- Fowler, D., Pitcairn, C.E.R., Sutton, M.a., Flechard, c., Loubet, B., Coyle, M. & Munro, R.C. (1998) The mass budget of atmospheric ammonia within 1km of livestock buildings. *Environ. Pollut.* (Nitrogen Conference Special Issue) **102**, S1:343-348.
- James, P.W., Hawksworth, D.H. & Rose, F. (1977) Lichen communities in the British Isles. In *Lichen Ecology*. ed. Seaward, M.R.D.: 295-413. Academic press, London.
- NEGTAP 2001. Transboundary Air Pollution (2001) DEFRA.

http://www.nbu.ac.uk/negtap/

- Nimis. PL, Scheidegger, C. and Wolseley, P.A. (2002) *Monitoring with Lichens Monitoring Lichens*. Nato Science Series, Kluwer, Dordrecht. 408 pp.
- Rose, F. (1992). Temperate forest management: its effects on bryophyte and lichen floras and habitats. In: *Bryophytes and lichens in a changing environment. eds.* Bates and Farmer: 211-233. Clarendon Press, Oxford.
- UK National Ammonia monitoring network <u>http://www.nbu.ac.uk/cara/UKNAMN/uknamn.htm</u>
- Van Herk, C.M. (1999) Mapping of ammonia pollution with epiphytic lichens in the Netherlands. *Lichenologist* **31**: 9-20
- Van Herk, C.M. (2002) Epiphytes on wayside trees as an indicator of eutrophication in the Netherlands. In *Monitoring with Lichens – Monitoring Lichens*. Eds Nimis. PL, Scheidegger, C. and Wolseley, P.A. Nato Science Series, Kluwer, Dordrecht. 285-290.
- Van Herk, C.M., Aptroot, A. & van Dobben, H.F. (2002) Long-term monitoring in the Netherlands suggests that lichens respond to global warming. *Lichenologist* 34: 141-154
- Wolseley, P.A. (2002) Using lichens on twigs to assess changes in ambient atmospheric conditions. In *Monitoring with Lichens – Monitoring Lichens*. Eds Nimis. PL, Scheidegger, C. and Wolseley, P.A. Nato Science Series, Kluwer, Dordrecht. 291-294.
- Wolseley, P. A. & Pryor, K.V. (1999) The potential of epiphytic twig communities on Quercus petraea in a Welsh woodland site (Tycanol) for evaluating environmental changes. *Lichenologist* **31**: 41-61.